

RESEARCH ARTICLE OPEN ACCESS

Increasing Prevalence of Plant-Fungal Symbiosis Across Two Centuries of Environmental Change

Joshua C. Fowler^{1,2}  | Jacob Moutouama^{1,3} | Tom E. X. Miller¹ 

¹Department of BioSciences, Rice University, Houston, Texas, USA | ²Department of Biology, University of Miami, Miami, Florida, USA | ³Department of Botany, University of British Columbia, Vancouver, British Columbia, Canada

Correspondence: Joshua C. Fowler (jcf221@miami.edu)

Received: 4 October 2024 | **Revised:** 6 October 2025 | **Accepted:** 6 October 2025

Funding: This research was supported by funding from the National Science Foundation Division of Environmental Biology (grants 1754468 and 2208857) and by funding from the Texas Ecolab Program. Five anonymous reviewers greatly improved earlier versions of this manuscript.

Keywords: climate change | *Epichloë* | herbarium | INLA | museum specimen | plant-microbe symbiosis | Poaceae | spatially varying coefficients model

ABSTRACT

Species' distributions and abundances are shifting in response to ongoing global climate change. Mutualistic microbial symbionts can provide hosts with protection from environmental stress that may promote resilience under environmental change; however, this change may also disrupt species interactions and lead to declines in hosts and/or symbionts. Symbionts preserved within natural history specimens offer a unique opportunity to quantify changes in microbial symbiosis across broad temporal and spatial scales. We asked how the prevalence of seed-transmitted fungal symbionts of grasses (*Epichloë* endophytes) has changed over time in response to climate change, and how these changes vary across host species' distributions. Specifically, we examined 2346 herbarium specimens of three grass host species (*Agrostis hyemalis*, *Agrostis perennans*, *Elymus virginicus*) collected over the past two centuries (1824–2019) for the presence or absence of *Epichloë* symbiosis. Analysis of an approximate Bayesian spatially varying coefficients model revealed that endophytes increased in prevalence over the last two centuries from ca. 25% to ca. 75% prevalence, on average, across three host species. Changes in seasonal climate drivers were associated with increasing endophyte prevalence. Notably, increasing precipitation during the peak growing season for *Agrostis* species and decreasing precipitation for *E. virginicus* were associated with increasing endophyte prevalence. Changes in the variability of precipitation and temperature during off-peak seasons were also important predictors of increasing endophyte prevalence. Our model performed favorably in an out-of-sample predictive test with contemporary survey data from across 63 populations, a rare extra step in collections-based research. However, there was greater local-scale variability in endophyte prevalence in contemporary data compared to model predictions, suggesting new directions that could improve predictive accuracy. Our results provide novel evidence for a cryptic biological response to climate change that may contribute to the resilience of host-microbe symbiosis through fitness benefits to symbiotic hosts.

1 | Introduction

Understanding how biotic interactions are altered by global change is a major goal of basic and applied ecological research (Blois et al. 2013; Gilman et al. 2010). Documented responses to environmental change, such as shifts in species' distributions

(Aitken et al. 2008) and phenology (Piao et al. 2019), are typically blind to concurrent changes in associated biotic interactions. Empirically evaluating these biotic changes—whether interacting species shift in tandem with their partners or not (HilleRisLambers et al. 2013)—is crucial to predicting the reorganization of Earth's biodiversity under global change. Such

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2025 The Author(s). *Global Change Biology* published by John Wiley & Sons Ltd.

evaluations have been limited because few datasets on species interactions extend over sufficiently long time scales of contemporary climate change (Poisot et al. 2021).

Natural history specimens, which were originally collected to document and preserve taxonomic diversity, present a unique opportunity to explore long-term changes in biodiversity and ecological interactions across broad spatial and temporal scales (Davis 2023; Meineke et al. 2018). Natural history collections, built and maintained by the efforts of thousands of scientists, are invaluable time machines, primarily comprised of physical specimens of organisms along with information about the time and place of their collection. These specimens often preserve physical legacies of ecological processes and species' interactions from dynamically changing environments across time and space (Lendemer et al. 2020). For example, previous researchers have examined the flowers, pollen grains, and leaves of specimens within plant collections (herbaria) to document shifts in reproductive phenology (Berg et al. 2019; Park et al. 2019; Willis et al. 2017), pollination (Duan et al. 2019; Pauw and Hawkins 2011), and herbivory (Meineke et al. 2019) related to anthropogenic climate change. Herbarium specimens have also been used to identify the origins and population genomics of plant diseases such as *Phytophthora*, the Irish potato famine pathogen (Ristaino et al. 2001; J. B. Ristaino 2002; Yoshida et al. 2013), and have been proposed as vehicles to track other emerging plant pathogens (Bradshaw et al. 2021; Jean B. Ristaino 2020). However, few previous studies have leveraged biological collections to examine climate change-related shifts in a particularly common type of interaction: mutualistic microbial symbiosis.

Microbial symbionts are common to all macroscopic organisms and can have important effects on their hosts' survival, growth, and reproduction (McFall-Ngai et al. 2013; Rodriguez et al. 2009). Many microbial symbionts act as mutualists, engaging in reciprocally beneficial interactions with their hosts in ways that can ameliorate environmental stress. For example, bacterial symbionts of insects, such as *Wolbachia*, can improve their hosts' thermal tolerance (Renoz et al. 2019; Truitt et al. 2019), and arbuscular mycorrhizal fungi, documented in 70%–90% of families of land plants (Parniske 2008), allow their hosts to persist through drought conditions by improving water and nutrient uptake (Cheng et al. 2021). On the other hand, changes in the mean and variance of environmental conditions may disrupt microbial mutualisms by changing the costs and benefits of the interaction for each partner in ways that can cause the interaction to deteriorate (Aslan et al. 2013; Fowler et al. 2024). Coral bleaching (the loss of symbiotic algae) due to temperature stress (Sully et al. 2019) is perhaps the best-known example, but this phenomenon is not unique to corals. Lichens exposed to elevated temperatures experienced loss of photosynthetic function along with changes in the composition of their algal symbiont community (Meyer et al. 2022). How commonly and under what conditions microbial mutualisms deteriorate or strengthen under climate change remain unanswered questions (Frederickson 2017). Previous work suggests that these alternative responses may depend on the intimacy and specialization of the interaction as well as the physiological tolerances of the mutualist partners (Rafferty et al. 2015; Toby Kiers et al. 2010; Warren and Bradford 2014).

Understanding how microbial symbioses are affected by climate change is additionally complicated by spatial heterogeneity in the direction and magnitude of environmental change (IPCC 2021). Beneficial symbionts are likely able to shield their hosts from environmental stress in locations that experience a small degree of change, but symbionts in locations that experience changes of large magnitude may be pushed beyond their physiological limits (Webster et al. 2008). Additionally, symbionts are often unevenly distributed across their host's distribution. Facultative symbionts may be absent from portions of the host range (Afkhami et al. 2014), and hosts may engage with a diversity of partners (different symbiont species or locally-adapted strains) across environments (Fowler et al. 2023; Frade et al. 2008; Rolshausen et al. 2018). Identifying broader spatial trends in symbiont prevalence is therefore an important step in developing predictions for where to expect changes in the symbiosis in future climates.

Epichloë fungal endophytes are specialized symbionts of cool-season grasses, estimated from surveys to associate with ~20%–30% of species across the diverse Poaceae family (Leuchtmann 1992). Within the cool-season grass subfamily (Pooideae), it has been estimated that between 17% to 40% of sampled species act as *Epichloë* hosts (Card et al. 2014; Iannone et al. 2011). They are predominantly transmitted vertically from maternal plants to offspring through seeds. Vertical transmission creates a feedback between the fitness of host and symbiont (Douglas 1998; Fine 1975; Rudgers et al. 2009). Over time, endophytes that act as mutualists should rise in prevalence within a host population, particularly under environmental conditions that elicit protective benefits (Donald et al. 2021). *Epichloë* are known to improve their hosts' drought tolerance (Decunta et al. 2021) and protect their hosts against herbivores (Ambrose et al. 2014; Crawford et al. 2010) and pathogens (Tian et al. 2017; Xia et al. 2018) likely through the production of a suite of biologically active molecules, including diverse alkaloids, proteins, and other secondary metabolites. The fitness feedback induced by vertical transmission leads to the prediction that endophyte prevalence should be high in populations where these fitness benefits are most important. Previous survey studies of contemporary populations have documented large-scale spatial patterns in endophyte prevalence structured by environmental gradients (Afkhami 2012; Bazely et al. 2007; Granath et al. 2007; Sneek et al. 2017). We predicted that endophyte prevalence should track temporal changes in environmental drivers (i.e., drought) that elicit strong fitness benefits.

Early research on *Epichloë* used herbarium specimens to describe the broad taxonomic diversity of grass host species that harbor these symbionts (White and Cole 1985), establishing that endophyte symbiosis could be identified in plant tissue from as early as 1851. However, no subsequent studies, to our knowledge, have used the vast resources of biological collections to quantitatively assess spatio-temporal trends in endophyte prevalence and their environmental correlates. Grasses are commonly collected and identified based on the presence of their reproductive structures, meaning that preserved specimens typically contain seeds, conveniently preserving the seed-transmitted fungi along with their host plants on herbarium sheets. This creates the opportunity to leverage the

unique spatio-temporal sampling of herbarium collections to examine the response of this symbiosis to historical climate change. However, the predictive ability derived from historical analyses is rarely tested against contemporary data (Lee et al. 2024). Critically evaluating whether insights from historical reconstruction are predictive of variation across contemporary populations is a crucial step for the field to move from reading signatures of the past to forecasting ecological dynamics into the future.

In this study, we assessed the long-term responses of *Epichloë* endophyte symbiosis to climate change through the use of herbarium specimens of three North American host grass species (*Agrostis hyemalis*, *Agrostis perennans*, and *Elymus virginicus*). We first addressed questions describing spatial and temporal trends in endophyte prevalence: (i) How has endophyte prevalence changed over the past two centuries? and (ii) How spatially variable are temporal trends in endophyte prevalence across eastern North America? We then addressed how climate change may be driving trends in endophyte prevalence by asking: (iii) What is the relationship between temporal trends in endophyte prevalence and associated changes in climate drivers? We predicted that overall endophyte prevalence would increase over time in tandem with climate change, and that localized hotspots of endophyte change would correspond spatially to hotspots of climate warming and drying. Finally, we evaluated (iv) how our model, built on data from historic specimens, performed in an out-of-sample test using data on endophyte prevalence from contemporary surveys of host populations. To answer these questions we examined a total of 2346 historic specimens collected across eastern North America between 1824 and 2019, and evaluated model performance against contemporary surveys comprising 1442 individuals from 63 populations surveyed between 2013 and 2020.

2 | Methods

2.1 | Focal Species

Our surveys focused on three native North American grasses: *Agrostis hyemalis*, *Agrostis perennans*, and *Elymus virginicus* that host *Epichloë* symbionts. These cool-season grass species have broad distributions covering much the eastern United States (Figure 1) and are commonly represented in natural history collections. Cool-season grasses grow during the cooler temperatures of spring and autumn due to their reliance on C_3 photosynthesis. *A. hyemalis* is a small short-lived perennial species that germinates in autumn to late winter and typically flowers between March and July (most common collection month: May). *A. perennans* is of similar stature but is longer lived than *Agrostis hyemalis* and flowers in late summer and early autumn (most common collection month: September). *A. perennans* is more sparsely distributed, tending to be found in shadier and moister habitats, while *A. hyemalis* is commonly found in open and recently disturbed habitats. Both *Agrostis* species are recorded from throughout the Eastern US, but *A. perennans* has a slightly more northern distribution, whereas *A. hyemalis* is found rarely as far north as Canada and is listed as a rare plant in Minnesota. *E. virginicus* is a larger and longer-lived species that is more broadly distributed than the *Agrostis* species. It begins flowering as early as March or April but continues throughout the summer (most common collection month: July).

Both *Agrostis* species host *Epichloë amarillans* (Craven et al. 2001; Leuchtman et al. 2014), and *Elymus virginicus* typically hosts *Epichloë elymi* (Clay and Schardl 2002). The fungal symbionts primarily reproduce asexually and are passed from maternal plant to offspring by vertical transmission through

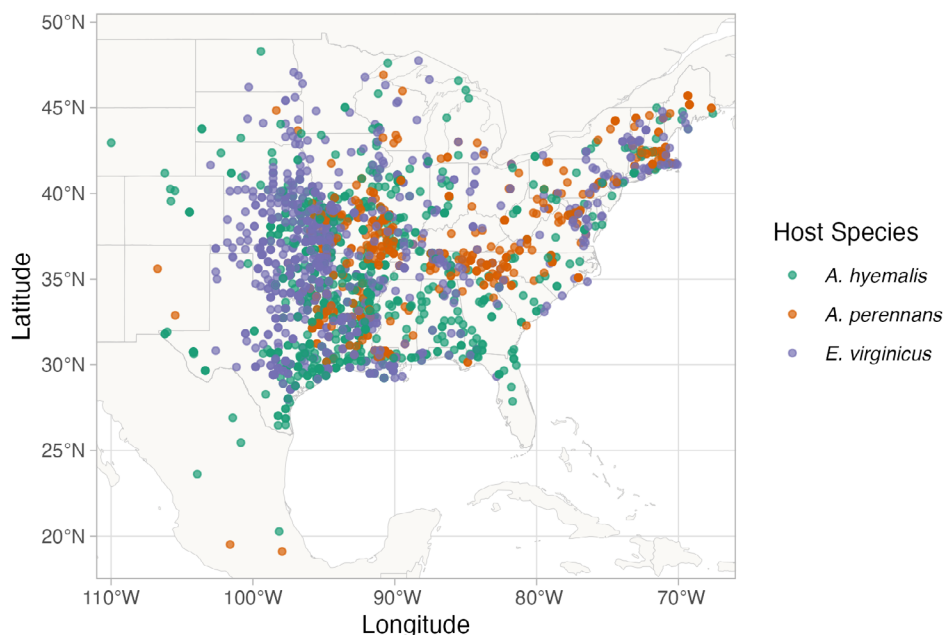


FIGURE 1 | Collection locations of herbarium specimens sampled for *Epichloë* endophytes. Specimens span eastern North America from nine herbaria, and are colored by host species (*A. hyemalis*: Green, *A. perennans*: Orange, *E. virginicus*: Purple). Map lines delineate study areas and do not necessarily depict accepted national boundaries.

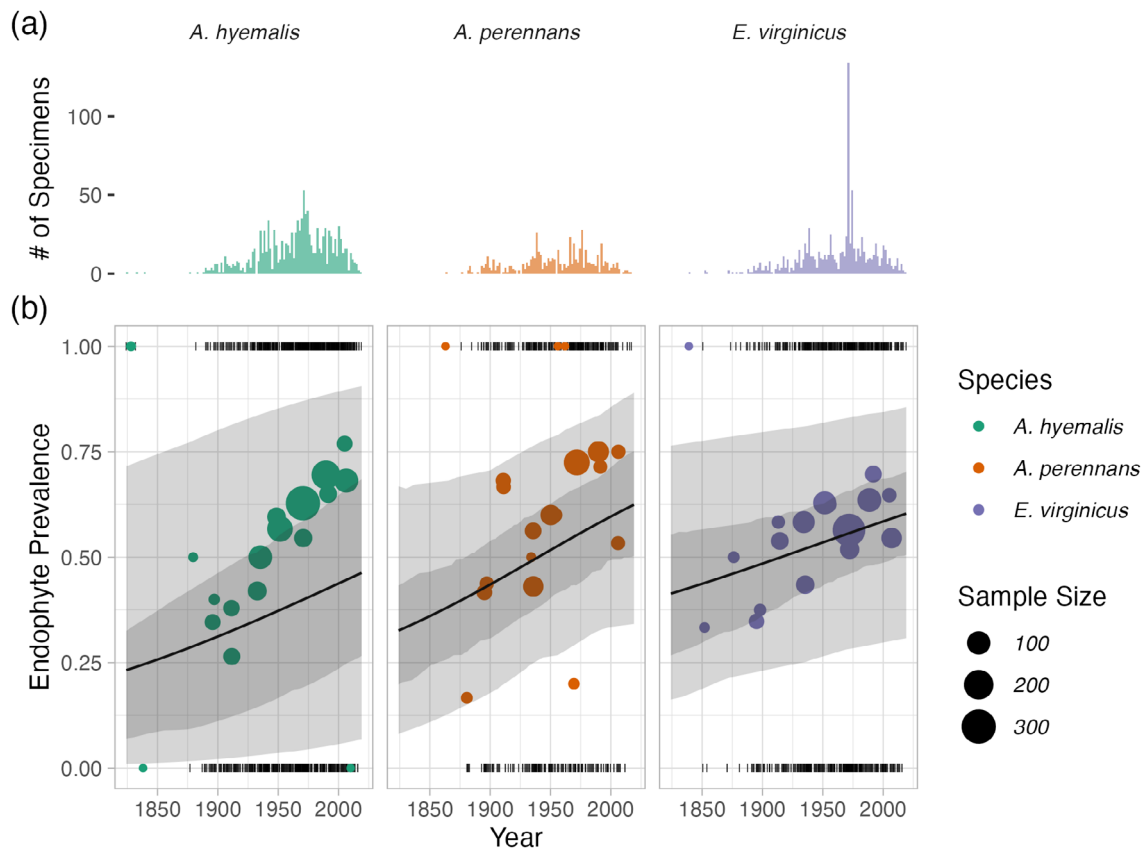


FIGURE 2 | Temporal trends in endophyte prevalence. (a) Histograms show the frequency of scored specimens through time for each host species. (b) Lines show mean endophyte prevalence predicted by the endophyte prevalence model over the study period along with the 50% and 95% CI bands incorporating parameter uncertainty and variation associated with collector and scorer random effects. Colored points are binned means of the observed endophyte presence/absence data (black dashes). Colors represent each host species (*A. hyemalis*: Green, *A. perennans*: Orange, *E. virginicus*: Purple) and point size represents the number of specimens.

seeds. These symbionts are also capable of horizontal transmission between hosts via the production of external reproductive structures, including sexual spore-bearing stromata that grow over host inflorescences (known as ‘choke disease’) and epiphyllous conidia on leaf surfaces that produce asexual spores (Tadych et al. 2014). Evidence suggests that the production of horizontal transmission structures by *Epichloë* occurs at low levels and may be influenced by environmental and genotypic factors (Brem and Leuchtman 1999; Meijer and Leuchtman 2000; Tintjer et al. 2008). In line with this, monitoring of long-term plots of *A. perennans* and *E. virginicus* showed no production of stromata on *A. perennans* and on only less than 1% of *E. virginicus* plants over 14 years (Fowler et al. 2024). A similar low frequency of stromata formation (only 0.37% of recorded inflorescences) was observed for *A. hyemalis* in a separate field experiment (Donald et al. 2021). Some host species have shown the capacity to partner with multiple symbiont species or strains, and in some cases, multiple symbiont lineages can coexist within a host population (Mc Cargo et al. 2014). However, surveys have typically found limited *Epichloë* genotypic diversity within host populations (Treindl et al. 2023). Across host populations, concentrations of biologically active biomolecules and the genes associated with their production vary substantially (Schardl et al. 2012). In this analysis, we focus on the presence/absence of *Epichloë* symbionts, and we discuss potential implications of symbiont genotypic diversity in the Discussion.

2.2 | Herbarium Surveys

We visited nine herbaria between 2019 and 2022 (see Table A1 for a summary of specimens included from each collection). With permission from herbarium staff, we acquired seed samples from 1135 *A. hyemalis* specimens collected between 1824 and 2019, 357 *A. perennans* specimens collected between 1863 and 2017, and 854 *E. virginicus* specimens collected between 1839 and 2019 (Figure 1, Figure 2a, Figure S1). We chose our focal species in part because they are commonly represented in herbarium collections and produce many seeds, meaning that small samples would not diminish the value of specimens for future studies. We collected 5–10 seeds per specimen after examining the herbarium sheet under a dissecting microscope to ensure that we collected mature seeds, not florets or unfilled seeds, fit for our purpose of identifying fungal endophytes with microscopy. We excluded specimens for which information about the collection location and date were unavailable.

Each specimen was assigned geographic coordinates based on collection information recorded on the herbarium sheet using the geocoding functionality of the *ggmap* R package (Kahle and Wickham 2019). Many specimens had digitized collection information readily available, but for those that did not, we transcribed information printed on the herbarium sheet. The identity of each specimen collector was gathered as part

of the sample's metadata. Collections were geo-referenced to the nearest county centroid, or nearest municipality when that information was available. For fifteen of the oldest specimens, only information at the state level was available, and so we used the state centroid. The median pairwise distance between georeferenced coordinate points was 841 km. The median longitudinal width of the bounding boxes generated to geocode municipality, county, or state centroids was 44.7 km. Among those specimens geo-referenced at the state level, the largest bounding box, spanning the state of Texas, was 1233 km wide. The smallest bounding boxes were less than 1 km across for small municipalities (while this suggests high precision, we note that some specimens were collected in natural habitat nearby to small municipalities not encompassed by these bounding boxes).

Our visits focused on herbaria with historic strengths in grass collections (e.g., Texas A&M, Missouri Botanic Garden) and other herbaria in the Southern Great Plains region of the United States. While these nine herbaria garnered specimens that span the focal species' ranges, our dataset unevenly samples across the study region (Figure 1). Texas, Oklahoma, Louisiana, and Missouri are the most represented states. Uneven sampling was most pronounced for *A. perennans*, which has much of its range in the northeastern US. We explore the potential influence of spatial bias in sampling on our results through a simulation analysis (Appendix S1—Supporting Methods).

After collecting seed samples, we quantified the presence or absence of *Epichloë* fungal hyphae in each specimen using microscopy. We first softened seeds with a 10% NaOH solution, then stained the seeds with aniline blue-lactic acid stain and squashed them under a microscope cover slip. We examined the squashed seeds for the presence of fungal hyphae at 200–400X magnification (Bacon and White 2018). On average we scored 4.7 intact seeds per specimen of *A. hyemalis*, 4.2 seeds per specimen of *A. perennans*, and 3.8 seeds per specimen of *E. virginicus*; we scored 10,342 seeds in total. Due to imperfect vertical transmission, the production of symbiont-free offspring from symbiotic hosts (Afkhani and Rudgers 2008), it is possible that symbiotic host-plants produce a mixture of symbiotic and nonsymbiotic seeds. We therefore designated a specimen as endophyte-symbiotic if *Epichloë* hyphae were observed in one or more of its seeds, or nonsymbiotic if *Epichloë* hyphae were observed in none of its seeds. To capture uncertainty in the endophyte identification process, we recorded both a “liberal” and a “conservative” endophyte score for each plant specimen. When we confidently identified endophytes within a specimen's seeds, we assigned matching liberal and conservative scores. When we identified potential endophytes with unusual morphology, low uptake of stain, or a small amount of fungal hyphae across the scored seeds, we recorded a positive identification for the liberal score and a negative identification for the conservative score. We recorded the identity of each scorer as part of the data collection process. 89% of scored plants had matching liberal and conservative scores, reflecting high confidence in endophyte status. The following analyses used the liberal status, however repeating all analyses with the conservative status yielded qualitatively similar results (Figure S8).

2.3 | Modeling Spatial and Temporal Changes in Endophyte Prevalence

We assessed spatial and temporal changes in endophyte prevalence across each host distribution, quantifying the “global” temporal trends averaged across space, and then examining spatial heterogeneity in the direction and magnitude of endophyte change (hotspots and coldspots) across the spatial extent of each host's distribution. To account for the spatial nonindependence of geo-referenced occurrences, we used an approximate Bayesian method, Integrated Nested Laplace Approximation (INLA), to construct spatio-temporal models of endophyte prevalence. INLA provides a computationally efficient method of ascertaining parameter posterior distributions for certain models that can be formulated as latent Gaussian Models (Rue et al. 2009). Many common statistical models, including structured and unstructured mixed-effects models, can be represented as latent Gaussian Models. We incorporated spatial heterogeneity into this analysis using spatially structured intercept and slope parameters implemented as stochastic partial differential equations (SPDE) to approximate a continuous spatial Gaussian process. This SPDE approach is a flexible method of smoothing across space while explicitly accounting for spatial dependence between data points (Bakka et al. 2018; Lindgren et al. 2011). Fitting models with structured spatial effects is possible with MCMC sampling but can require long computation times, making INLA an effective alternative. This approach has been used to model spatial patterns in flowering phenology (Willems et al. 2022), the abundance of birds (Meehan et al. 2019) and butterflies (Crossley et al. 2022), the distribution of temperate trees (Engel et al. 2022) as well as the population dynamics of endangered amphibians (Knapp et al. 2016) and other ecological processes (Beguín et al. 2012).

We estimated global and spatially-varying trends in endophyte prevalence using a joint-likelihood model. For each host species h , endophyte presence/absence of the i^{th} specimen ($P_{h,i}$) was modeled as a Bernoulli response variable with expected probability of endophyte occurrence $\hat{P}_{h,i}$. We modeled $\hat{P}_{h,i}$ as a linear function of collection year, with intercept A_h and slope T_h defining the global temporal trend in endophyte prevalence specific to each host species as well as with spatially-varying intercepts α_{h,l_i} and slopes τ_{h,l_i} associated with location (l_i , the unique latitude-longitude combination of the i^{th} observation). The joint-model structure allowed us to “borrow information” across species in the estimation of shared variance terms for the spatially-dependent random effect δ_{l_i} , intended to account for residual spatial variation, and χ_{c_i} and ω_{s_i} , the i.i.d.-random effects indexed for each collector identity (c_i) and scorer identity (s_i) of the i^{th} specimen.

$$\text{logit}(\hat{P}_{h,i}) = A_h + T_h * \text{year}_i + \alpha_{h,l_i} + \tau_{h,l_i} * \text{year}_i + \delta_{l_i} + \chi_{c_i} + \omega_{s_i} \quad (1)$$

By including random effects for collectors and scorers, we accounted for “nuisance” variance that may bias predictions for changes in endophyte prevalence. Previous work suggests that the behavior of historical botanists may introduce biases into ecological inferences made from historic collections (Kozlov et al. 2020). Prolific collectors who contribute thousands of

specimens may be more or less likely to collect certain species, or specimens with certain traits (Daru et al. 2018). Similarly, the process of scoring seeds for hyphae involved multiple researchers (or “scorers”) who, even with standardized training, may vary in their likelihood of positively identifying *Epichloë*.

We performed model fitting using the *inlabru* R package (Bachl et al. 2019). Global intercept and slope parameters, A and T , were given vague priors. Collector and scorer random effects, χ and ω respectively, were centered at 0 with precision parameters assigned penalized complexity (PC) priors with parameter values $U_{PC} = 1$ and $a_{PC} = 0.01$ (Simpson et al. 2017). Each spatially-structured parameter depended on a covariance matrix according to the proximity of each pair of collection locations (Bakka et al. 2018; Lindgren et al. 2011). The covariance matrix was approximated using a Matérn covariance function, with each data point assigned a location according to the nodes of a mesh of nonoverlapping triangles encompassing the study area (Figure S2). Matérn covariance functions are widely used in spatially explicit statistical modeling because of their mathematical tractability and flexibility. This covariance structure relies on the assumption that the underlying process is stationary and isotropic, such that spatial autocorrelation between data points depends only on their relative positions (Bakka et al. 2018).

Implementing spatially-structured parameters in INLA with this SPDE approach is useful particularly because space is treated as a continuous variable, allowing the model to make efficient use of the data and generate predictions across the entire study region. The SPDE approach is flexible enough that it can capture smooth trends across space that are informed by the data rather than by spatial regions chosen a priori by researchers. However this flexibility also invites the risk of overfitting, as with other nonlinear modeling approaches (Lapeyrolerie and Boettiger 2023; Ramampandra et al. 2023; Ward et al. 2014). Priors for the Matérn covariance function, termed “range” and “variance”, define how proximity effects decay with distance. The choice of priors for these types of spatial models is an area of active research (Bakka et al. 2018; Simpson et al. 2017), but another advantage of the INLA approach is that its computational efficiency allows for prior sensitivity analyses. Results presented in the main text reflect a prior range of 342 kilometers (i.e., a 50% probability of estimating a range less than 342 kilometers). We tested a range of values (from 68 kilometers to 1714 kilometers) and meshes (presented in the Supporting Methods—*Mesh and Prior Sensitivity Analysis*), finding that while the magnitude and uncertainty of effects varied, model results were qualitatively similar, that is, the same direction of effects across space. We assessed model fit with visual posterior predictive checks (Figure S3) and measurements of AUC (Figures S4 and S5) (Gelman and Hill 2006). Through results and discussion that follow, we refer to the model described in this section as the “endophyte prevalence model”.

2.4 | Modeling Distributions of Host Species

The herbarium records did not encompass the entirety of each host species’ range. Therefore, we used additional data sources to model the geographic distribution of each host species, with two goals: (1) generate realistic maps on which we could project the

predictions of the INLA model, and (2) use the geographic distributions to test for relationships between climate change drivers and trends in endophyte prevalence. We followed the ODMAP (overview, data, model, assessment, prediction) protocol (Crossley et al. 2022) (see Supporting Methods). In short, we used presence-only observations of each host species from the Global Biodiversity Information Facility (GBIF) between 1990 and 2020 (713 occurrence records for *A. hyemalis* (GBIF.Org 2025a), 656 occurrence records for *A. perennans* (GBIF.Org 2025b), and 2338 occurrence records for *E. virginicus* (GBIF.Org 2025c)). We fit maximum entropy (MaxEnt) models using the maxent function in the R package *dismo* (Hijmans et al. 2017) using the following seasonal climate predictors (1990–2020 climate normals): mean and standard deviation of spring, summer, and autumn temperature, and mean and standard deviation of spring, summer, and autumn cumulative precipitation.

We generated 10,000 pseudo-absences as background points, and split the occurrence data into 75% for model training and 25% for model testing. The performance of models was evaluated with AUC (Jiménez-Valverde 2012). We found AUC values of 0.862, 0.838, and 0.821, respectively for *Agrostis hyemalis*, *Agrostis perennans*, and *Elymus virginicus*, indicating good model fit to data. We used the training sensitivity (true positive rate) and specificity (true negative rate) to set a threshold for transforming the continuous predicted probabilities into binary presence—absence host distribution maps on which we projected INLA predictions of endophyte prevalence (Liu et al. 2005).

2.5 | Assessing the Role of Climate Drivers

We assessed how the magnitude of climate change may have driven changes in endophyte prevalence by assessing correlations between changes in climate and changes in endophyte prevalence predicted from our spatial model at evenly spaced pixels across the study area.

We first downloaded monthly temperature and precipitation rasters from the PRISM climate group (Daly and Bryant 2013) covering the time period between 1895 and 2020 using the *prism* R package (Hart and Bell 2015). PRISM provides reconstructions of historic climate variables across the United States by spatially interpolating weather station data (Di Luzio et al. 2008). Because the magnitude of observed climate change differs across seasons, and because different growing seasons are a key feature of the biology of our focal host species, we calculated 30-year climate normals for seasonal mean temperature and cumulative precipitation for the recent (1990 to 2020) and historic (1895 to 1925) periods. We used three four-month seasons within the year (Spring: January, February, March, April; Summer: May, June, July, August; Autumn: September, October, November, December). This division of seasons allowed us to quantify differences in the primary climate change drivers, temperature and precipitation, associated with the two “cool” seasons, when we expected our focal species to be most active (*A. hyemalis* flowering phenology: spring; *E. virginicus*: spring and summer; *A. perennans*: autumn). In addition to mean climate conditions, environmental variability itself can influence population dynamics (Tuljapurkar 1982) and changes in variability are a key prediction of climate change models (IPCC 2021; Stocker et al. 2013). Therefore, we calculated the

standard deviation for each annual and seasonal climate driver across each 30-year period. We then took the difference between recent and historic periods for the mean and standard deviation for each climate driver (Figures S13–S15). All together, we assessed twelve potential climate drivers: the mean and standard deviation of spring, summer, and autumn temperature, as well as the mean and standard deviation of spring, summer, and autumn cumulative precipitation (the same climate covariates used in the MaxEnt models).

We then evaluated whether areas that have experienced the greatest changes in endophyte prevalence (hotspots of endophyte change) are associated with high degrees of change in climate (hotspots of climate change). To do so, we modeled the fitted, spatially-varying slopes of endophyte change through time ($\tau_{h,t}$) as a linear function of environmental covariates, with a Gaussian error distribution for a set of pixels across each host distribution. The continuous SPDE approach taken from our endophyte prevalence model allows us to generate predictions of temporal trends in prevalence at arbitrarily many pixels across each host distribution. Balancing computation time with resolution, we generated predicted trends for 546, 645, and 753 pixels across each host distribution for *A. perennans*, *A. hyemalis*, and *E. virginicus* respectively (pixel dimensions: *A. perennans* = 65 km x 36 km; *A. hyemalis* = 61 km x 45 km; *E. virginicus* = 62 km x 40 km). Fitting regressions to many pixels across the study region risks artificially inflating confidence in our results due to large sample sizes, and so we performed this analysis using only a random subsample of 250 pixels across the study region; other sizes of subsample yielded similar results. Data from each host species were analyzed separately. Throughout the results and discussion that follow, we refer to this analysis as the “*post hoc* climate regression analysis”.

2.6 | Validating Model Performance With In-Sample and Out-Of-Sample Tests

We evaluated the predictive ability of the endophyte prevalence model using both in-sample training data from the herbarium surveys, and with out-of-sample test data, an important but rarely used strategy in ecological studies (Lee et al. 2024; Tredennick et al. 2021). We generated out-of-sample test data from contemporary surveys of endophyte prevalence in natural populations of *A. hyemalis* and *E. virginicus* in Texas and the southern US. Surveys of *E. virginicus* were conducted in 2013 as described in Sneek et al. (2017), and surveys of *A. hyemalis* took place between 2015 and 2020. Population surveys of *A. hyemalis* were initially designed to cover longitudinal variation in endophyte prevalence towards its range edge, while surveys of *E. virginicus* were designed to cover latitudinal variation. In total, we visited 43 populations of *A. hyemalis* and 20 populations of *E. virginicus* across the south-central US, with emphasis on Texas and neighboring states (Figure S12). Number of plants sampled per population: 22.9; note that this sampling design provided greater local depth of information than the herbarium records, where only one plant was sampled at each locality. We quantified the endophyte status of each individual with microscopy as described for the herbarium surveys (with 5–10 seeds scored per individual), and calculated the prevalence of endophytes within the population (proportion of plants that

were endophyte-symbiotic). For each population, we compared the observed fraction of endophyte-symbiotic hosts to the predicted probability of endophyte occurrence \hat{P} derived from the model for that location and year. The contemporary survey period (2013–2020) is at the most recent edge of the time period encompassed by the historical specimens used for model fitting.

3 | Results

3.1 | How Has Endophyte Prevalence Changed Over Time?

Across more than 2300 herbarium specimens dating back to 1824, we found that prevalence of *Epichloë* endophytes increased over the last two centuries for all three grass host species (Figure 2). On average, endophytes of *A. perennans* and *E. virginicus* increased from ~40% to 70% prevalence across the study region, and *A. hyemalis* increased from ~25% to over 50% prevalence. Our model indicates high confidence that overall temporal trends are positive across species (99% probability of a positive overall year slope in *A. hyemalis*, 92% probability of a positive overall year slope in *A. perennans*, and 91% probability of a positive overall year slope in *E. virginicus*) (Figure S6).

The model appears to under-predict the observed increase in endophyte prevalence relative to the data, particularly for *A. hyemalis* (Figure 2b), but the model is accounting for random effects and spatial nonindependence that are not readily seen in the figure. We found no evidence that collector biases influenced our results. Collector random effects were consistently small (Figure S9), and models fit with and without this random effect provide qualitatively similar results. The identity of individual scorers, the researchers who identified endophyte status microscopically, did contribute to observed patterns in endophyte prevalence. For example, 3 of the 25 scorers were significantly more likely than average to assign positive endophyte status, as indicated by 95% credible intervals greater than zero, while 4 of the 25 had 95% credible intervals below zero (Figure S10).

3.2 | How Spatially Variable Are Temporal Trends in Endophyte Prevalence?

While there was an overall increase in endophyte prevalence, our model revealed hotspots and coldspots of change across the host species' ranges, which are mapped in Figure 3 across geographic ranges predicted by MaxEnt species distribution models. In some regions, posterior mean estimates of spatially varying temporal trends indicate that *A. hyemalis* and *A. perennans* experienced increases in prevalence by as much as 2% per year over the study period. Posterior estimates of uncertainty in spatially varying slopes indicate that these hotspots of change may have experienced increases of up to 5% per year while declines in prevalence may be as great as –4% per year for the *Agrostis* species. (Figure S7) In contrast, *E. virginicus* experienced increases up to around 1% per year, with uncertainty ranging between 3.5% increases and 2.5% decreases (Figure S7) Taken together, both *Agrostis* species show areas of both strong increasing and declining prevalence, while *E. virginicus* had an overall weaker and geographically more homogeneous increase

in endophyte prevalence. Notably, endophytes are predicted to have increased most strongly towards the western range edge of *A. hyemalis* (Figure 3a) and across the northeastern US for *A. perennans* (Figure 3b). Broad increases in prevalence on average, along with increases towards range edges that had low historic prevalence result in range expansions of the symbiosis for both *Agrostis* species (Figure 4). Increases in prevalence were strongest in regions with low historic prevalence for the *Agrostis* species (Figure S11A,B), but for *E. virginicus* trends did not differ according to historic prevalence (A11 C).

3.3 | What Is the Relationship Between Variation in Temporal Trends in Endophyte Prevalence and Changes in Climate Drivers?

We found that trends in endophyte prevalence were strongly associated with one or more seasonal climate change drivers (Figure 5). For the majority of the study region, the climate has become wetter (an average increase in annual precipitation of 60 mm) with relatively minimal temperature warming (an average increase in annual temperature of 0.02°C) over the last

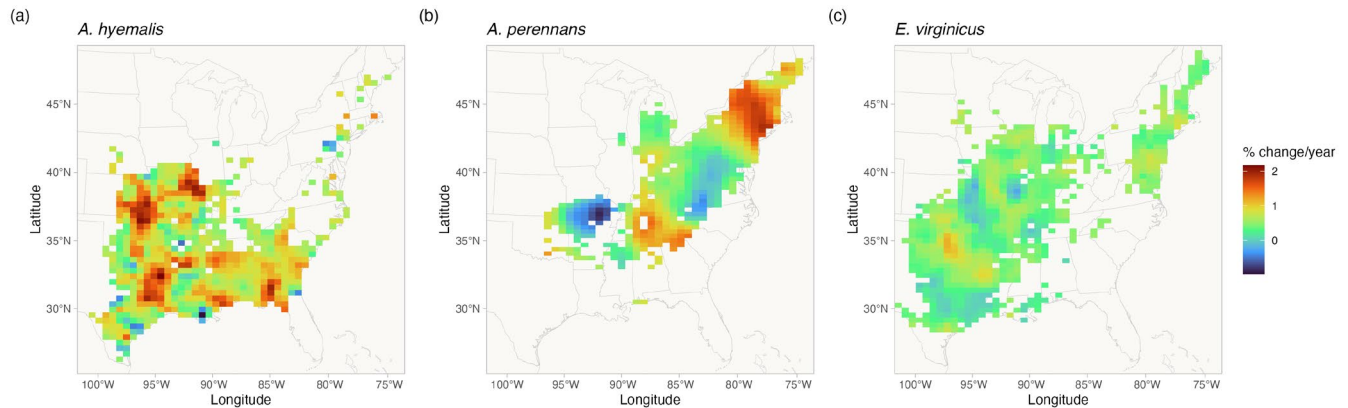


FIGURE 3 | Predicted posterior mean of spatially varying slopes representing change in endophyte prevalence for each host species ((a) *A. hyemalis*; (b) *A. perennans*; (c) *E. virginicus*). Spatially varying trends are estimated from the endophyte prevalence model. Color indicates the relative change in predicted endophyte prevalence. Map lines delineate study areas and do not necessarily depict accepted national boundaries.

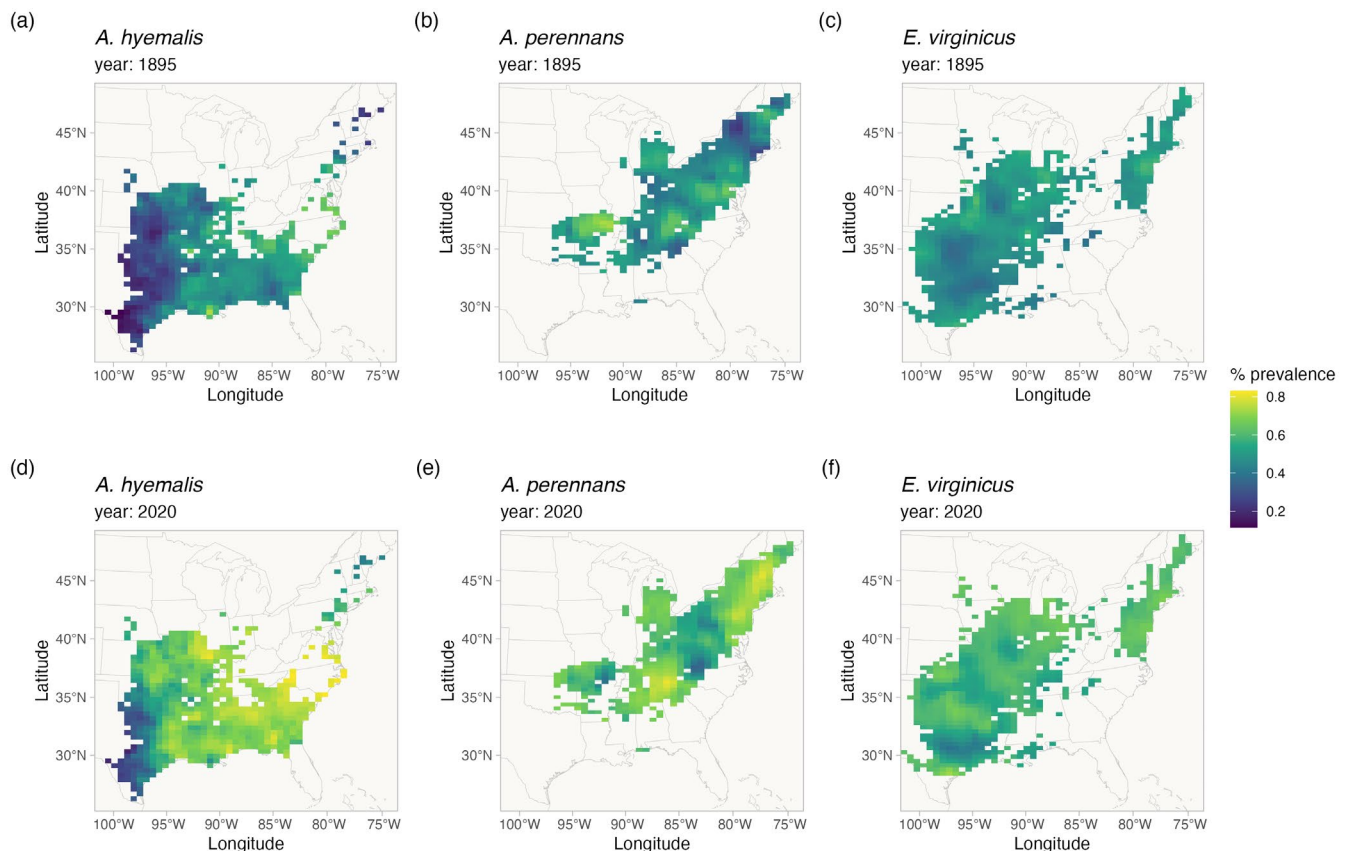


FIGURE 4 | Predicted endophyte prevalence for each host species in 1895 and 2020. Predictions of prevalence come from the endophyte prevalence model. Color indicates the posterior mean endophyte prevalence for (a, d) *A. hyemalis*, (b, e) *A. perennans*, and (c, f) *E. virginicus*. Map lines delineate study areas and do not necessarily depict accepted national boundaries.

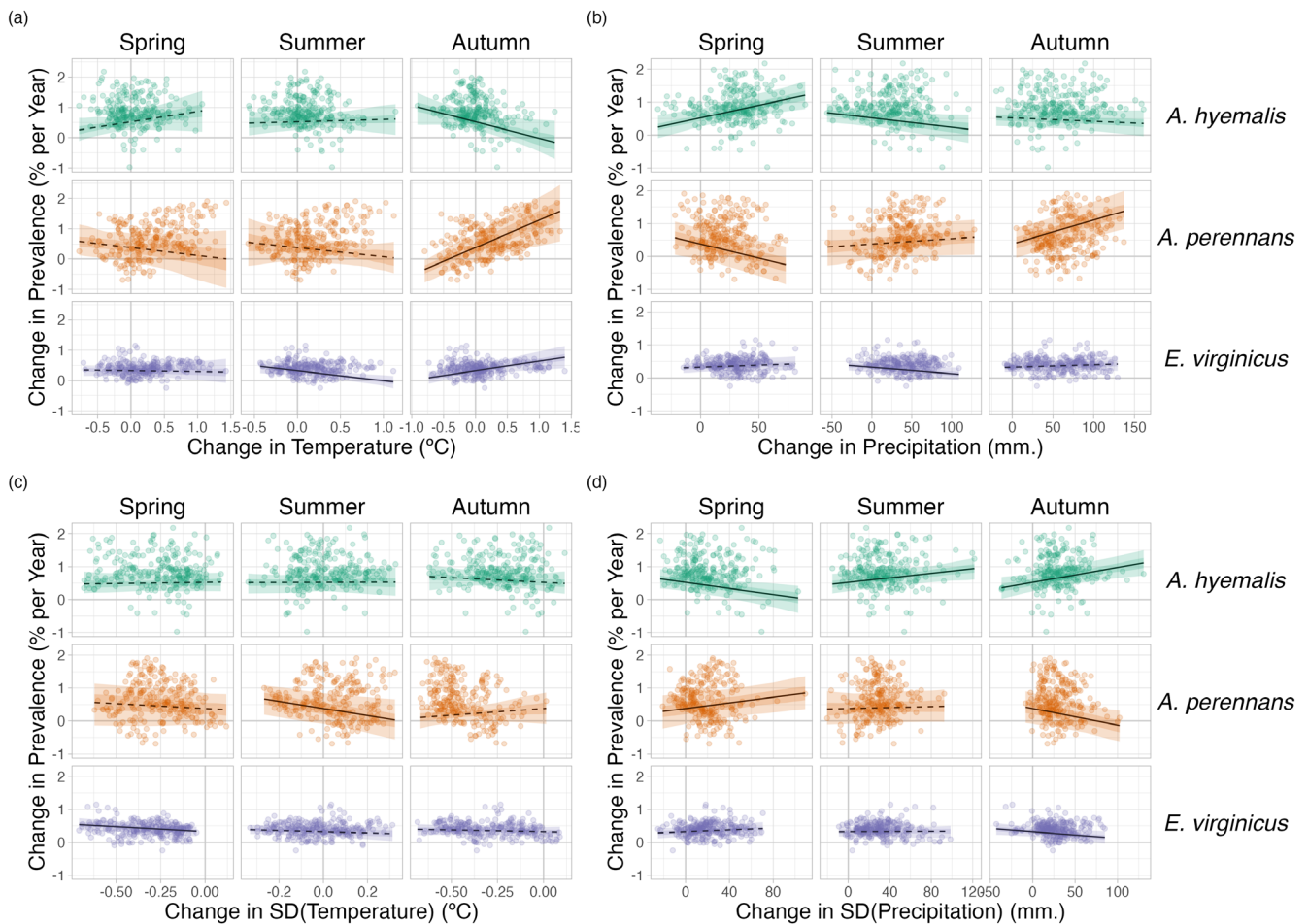


FIGURE 5 | Relationships between predicted trends in endophyte prevalence and changes in seasonal climate drivers. Lines show marginal predicted relationship between spatially-varying trends in endophyte prevalence and changes in mean and variability of climate ((a): Mean temperature, (b): Cumulative precipitation, (c): Standard deviation in temperature, (d): Standard deviation in precipitation) estimated from the *post hoc* climate regression analysis. Confidence bands represent the 50% and 95% CI, colored by host species (*A. hyemalis*: Green, *A. perennans*: Orange, *E. virginicus*: Purple). Slopes with greater than 95% posterior probability of being either positive or negative are represented as solid lines while those that have less than 95% probability are dashed. Points are the values of precomputed SVC trends and climate drivers at 250 randomly sampled pixels across each host's distribution used in model fitting for the *post hoc* climate regression analysis.

century (Figures S13–S15), a consequence of regional variation in global climate change (IPCC 2021). Within the region, climate changes were spatially variable; certain locations experienced increases in annual precipitation as large as 375 mm or decreases up to 54 mm across the last century, while annual temperature changes ranged from warming as great as 1.4°C to cooling by 0.46°C.

Spatially variable climate trends were predictive of trends in endophyte prevalence. For example, among the tested climate drivers, strong increases in endophyte prevalence for *A. perennans* were most strongly associated with increasing autumn precipitation and with increasing mean and variability in autumn temperature (greater than 97% posterior probabilities of positive slopes). For this species, each 1°C increase in autumn temperature was associated with a 1.07% greater increase per year in endophyte prevalence (Figure 5a) and a 100 mm increase in precipitation was associated with a 0.8% greater increase per year in endophyte prevalence (Figure 5b). This result aligns with the species' autumn active growing season, however other

seasonal climate drivers were also positively associated with increasing endophyte prevalence in this host species. In particular, we found cooler and drier springs and cooler summers to be associated with increasing endophyte prevalence (greater than 99% posterior probabilities of negative slopes), though these slopes were generally of smaller magnitude than those for autumn climate drivers. Changes in endophyte prevalence across the ranges of *A. hyemalis* and *E. virginicus* were less strongly driven by changes in climate. Like *A. perennans*, climate during peak growing season (spring for *A. perennans* and summer for *E. virginicus*) emerged most commonly as drivers of changes in endophyte prevalence. Across the tested climate drivers, increases in mean spring precipitation were the strongest predictor of increasing trends in endophyte prevalence for *A. hyemalis* (Figure 5b) (greater than 99% posterior probability of a positive slope). For this species, an increase of 100 mm in spring precipitation was associated with 0.6% per year stronger increases in endophyte prevalence relative to regions with no change in precipitation. The next greatest slopes were those associated with variability in spring precipitation (greater than 96% posterior

probability of a negative slope), as well as in the mean and variability of autumn climate (greater than 98% probability of negative and positive slopes, respectively). Changes in endophyte prevalence in *E. virginicus* were not strongly associated with changes in most climate drivers, but regions with reduced variability in autumn precipitation (Figure 5b) and with cooler and more variable summer temperatures (Figure 5a,c) experienced the largest increases in endophyte prevalence. Our analysis indicated relatively high confidence that these climate drivers influence endophyte prevalence shifts in *E. virginicus* (greater than 99% posterior probability of either negative or positive slopes respectively), however they translate, for example, to less than a 0.4% decrease in endophyte prevalence per year for each 1°C of summer warming over the century. Repeating this analysis using all pixels across each species' distribution were qualitatively similar to these results.

3.4 | Evaluation of Model Performance on an Out-Of-Sample Test

Tests of the endophyte prevalence model's predictive performance, as quantified by AUC and by visual posterior predictive checks, indicated good predictive ability. Model performance was similar between historic herbarium specimens used as training data and the out-of-sample test data from contemporary surveys (AUC=0.79 and 0.77 respectively; Figures S4 and S5). The model successfully captured broad regional trends in endophyte prevalence seen in the contemporary survey data, such as a decline in endophyte prevalence in *A. hyemalis* towards western longitudes (Figure 6a) and an increase towards northern latitudes (Figure 6b). It is notable that model predictions for endophyte prevalence exhibited relatively little local geographic variation, whereas the out-of-sample survey data were highly variable with

populations spanning 0% to 100% endophyte-symbiotic plants (Figure 6c), indicating population-to-population variation not captured in the endophyte prevalence model.

4 | Discussion

Our examination of historic plant specimens revealed previously hidden shifts in microbial symbiosis over the last two centuries. For the three grass host species we examined, there have been strong increases in the prevalence of *Epichloë* endophyte symbiosis. We interpret increases in the prevalence of *Epichloë*, which are predominantly vertically transmitted, as adaptive changes that improve the fitness of their hosts under increasing environmental stress. This interpretation is in line with theory predicting that positive fitness feedback caused by vertical transmission leads beneficial symbionts to rise in prevalence within a population (Donald et al. 2021; Fine 1975). We further found that trends in endophyte prevalence often varied across the host distribution in association with changes in climate drivers, consistent with the hypothesis that increases in endophyte prevalence are driven by context-dependent benefits to hosts that confer resilience under environmental change. Taken together, our results suggest an overall strengthening of host-symbiont mutualism over the last two centuries.

4.1 | Responses of Host-Microbe Symbioses to Climate Change

Differences across host species underscore that while all of these C_3 grasses share similar broad-scale distributions, each engages in unique biotic interactions and has unique responses to environmental drivers. We identified hotspots of change for

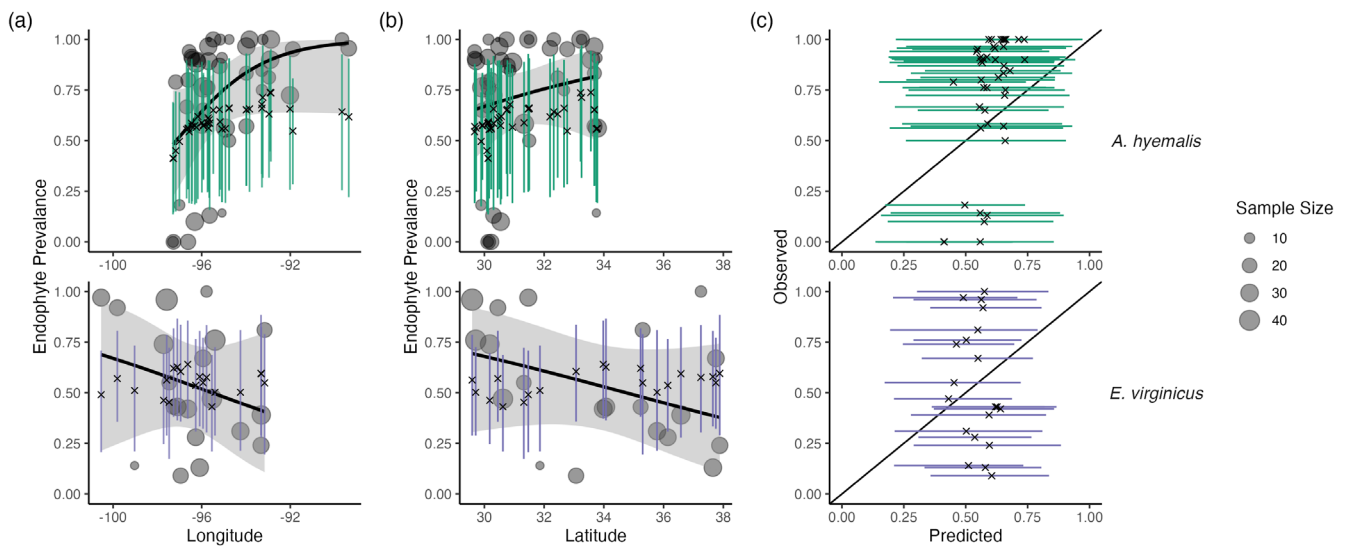


FIGURE 6 | Predictive performance for contemporary test data. (a) The endophyte prevalence model, trained on historic herbarium collection data, performed modestly at predicting prevalence in contemporary population surveys. The model captured regional trends across (a) longitude and (b) latitude. Crosses indicate predicted mean prevalence along with the 95% CI (colored lines: *A. hyemalis*: Green, orange, *E. virginicus*: Purple) from the herbarium model. Contemporary prevalence is represented by grey points (point size reflects sample size) along with trend lines from generalized linear models (black line and shaded 95% confidence interval). (c) Comparison of contemporary observed population prevalence vs. predicted endophyte prevalence shows that contemporary test data had more variance between populations than in model predictions.

A. perennans, which was the species whose endophyte prevalence was most responsive to changes in climate drivers (Figure 5). Predicted declines of 0.9% per year in the southern portion of its range and predicted increases of up to 2% per year in the north suggest a potential poleward range shift of endophyte-symbiotic plants (Figure 3b); whether the overall host distribution is shifting in parallel is an exciting next question.

Based on previous work demonstrating that endophytes can shield their hosts from drought stress (reviewed in Decunta et al. (2021)), we generally predicted that drought conditions would be a driver of increasing endophyte prevalence. In contrast to this expectation, increasing prevalence for *A. perennans* was associated with both increasing autumn temperature and precipitation (Figure 5). To our knowledge, the response of the symbiosis in *A. perennans* to drought has not been examined experimentally, but in a greenhouse experiment, endophytes had a positive effect on host reproduction under shaded, low-light conditions (Davitt et al. 2010). Our results also hint that it may be useful to investigate whether lagged climate effects are important predictors of host fitness in this system (Evers et al. 2021). Endophyte prevalence of the autumn-flowering *A. perennans* was strongly linked with decreasing spring precipitation, and that of the spring-flowering *A. hyemalis* was associated with decreasing autumn precipitation (Figure 5b). For *A. hyemalis*, endophytes could be playing a role helping hosts weather autumn-season droughts, which is likely also an important time for the species' germination. Previous work demonstrated drought benefits in a greenhouse manipulation with this host-symbiont pair (Davitt et al. 2011), and early life stages may be particularly vulnerable to prolonged droughts. For *E. virginicus*, which experienced the weakest changes in endophyte prevalence overall (ranging between 1.1% increases and 0.2% decreases), we only found modest associations with changes in climate drivers. Surveys by Sneek et al. (2017), used as part of the test data in this study, identified a drought index (SPEI) that integrates precipitation with estimated evapotranspiration as an important predictor of contemporary endophyte prevalence in this species. The diverse relationships we detect between trends in endophyte prevalence and climate drivers suggest a more complicated picture than the simple explanation that drought alone, as measured through changes in annual precipitation, causes increasing endophyte prevalence through context-dependent fitness benefits.

While we show consistent increasing trends in prevalence between the three species, the mechanisms that explain these changes may be diverse and idiosyncratic. First, climate change responses may depend on genotype-specific responses that are not considered in our current analysis. While *Epichloë* symbioses are highly specialized, surveys have demonstrated genotypic and chemotypic diversity of the symbionts among and within populations (Treindl et al. 2023; von Cräutlein et al. 2021). Genotypic variation in *Epichloë* endophytes, particularly in genes responsible for alkaloid production, produces "chemotypes" with differing benefits for hosts against insect or mammalian herbivores mediated by environmental conditions (Ambrose et al. 2014; Saikkonen et al. 2013; Schardl et al. 2012). Genotypic variation of the hosts themselves can also influence interaction outcomes (Gundel, Zabalgoeazcoa, and De Aldana 2011; Parker et al. 2017). Whether hotspots of change

in endophyte prevalence reflect selection for genotype pairings with particularly strong fitness benefits is an unanswered question. Additionally, *Epichloë* endophytes have been connected to a suite of nondrought-related fitness benefits including herbivory defense (Brem and Leuchtman 2001), salinity resistance (Wang et al. 2020), and mediation of pathogens (Vikuk et al. 2019) and the soil microbiome (Roberts and Ferraro 2015). Broad changes in the distribution and abundance of natural enemies (Côté et al. 2004), along with stresses from anthropogenic changes in land management and pollution (Sage 2020), likely influence the benefits of symbiosis (Rudgers et al. 2020). Changing endophyte prevalence results from the combination of net fitness benefits playing out across the heterogeneous map of a changing climate and its interactive effects on other anthropogenic drivers. Host species experience a world that is made increasingly stressful by a combination of global change drivers, and while historic trends that we observed suggest that symbiotic fitness benefits have helped mitigate this stress, it is possible that at yet higher levels of stress, increasing costs of the mutualism could lead to declines in endophyte prevalence. It is also known that stressful conditions can both (Gundel, Garibaldi, et al. 2011) and increase (Gundel et al. 2020) the rate of successful transmission of *Epichloë* endophytes from mother plant to offspring. These responses likely reflect both consequences of host sanctions on costly symbionts and the ability of symbionts to successfully colonize seeds (Afkhami and Rudgers 2008; Gundel, Rudgers, and Ghersa 2011). While we did not investigate differences in the rate of transmission across these historic specimens due to low numbers of sampled seeds per individual, it would be valuable to conduct deeper sampling for a subset of specimens with known endophyte status and investigate how transmission itself may respond to environmental change. Additionally, we have interpreted increasing prevalence as a signature of increased host fitness driven by vertical transmission, which we expect is the predominant mode of transmission in this system. Horizontal transmission, while rare, could also explain increasing prevalence if global change stressors lead symbionts to engage in increased parasitic behavior, resulting in increased dispersal and transmission at the cost of host fitness. More extreme climate stresses, which are expected more frequently in the future (Seneviratne et al. 2021), could shift the balance of interaction costs and benefits. Identifying "tipping points" of mutualism breakdown under increasing environmental stress is an important area of future inquiry.

Our results indicate that *Epichloë* symbiosis has likely improved host fitness in stressful environments leading to increasing prevalence. What is less clear is how this will influence future range shifts. Based on our analysis, it is likely that the symbiosis will facilitate range shifts for hosts by improving population growth at range edges. Previous population surveys (Rudgers and Swafford 2009; Semmartin et al. 2015; Sneek et al. 2017) attributed environment-dependent gradients in endophyte prevalence to symbiont-derived fitness benefits allowing hosts to persist in environments where they otherwise could not (Afkhami et al. 2014; Kazenel et al. 2015). However, symbiont-facilitated range shifts require that endophytes be present in the populations to be able to contribute to population growth. For example, the arid western range edge of *A. hyemalis* has had historically low endophyte prevalence (Figure 4), and dispersal of symbionts may limit the capacity for range shifts. A range

edge population with no endophytes would require dispersal of symbionts to occur, either via symbiotic seeds or horizontally-transmitted fungal spores, before the symbiosis could result in an expanding distribution (Fowler et al. 2023). At the same time, we found that endophyte prevalence has increased most quickly in regions with historically low endophyte prevalence (Figure S11), suggesting strong selection for symbiotic hosts when they are present. These factors potentially contribute to the ability of the host species to track its environmental niche. Another interesting question is the degree to which symbiotic and nonsymbiotic hosts, which occupy overlapping but distinct niches, are likely to experience distribution shifts in tandem or at different rates in the future.

4.2 | Steps Towards Forecasts of Host-Microbe Symbioses

The combination of a spatially explicit model and historic herbarium specimens allowed us to identify regions of both increasing and decreasing endophyte prevalence. We see several next steps toward the goal of predicting host and symbiont niche shifts in response to future climate change. While the model successfully predicted large-scale spatial trends observed in the out-of-sample contemporary population surveys, these data contained more population-to-population variability in prevalence than could be explained by the model. We interpret this to mean that the model captures coarse-scale spatial and temporal trends reasonably well, but is not equipped to capture local-scale nuances that generate population-to-population differences. Validating our model predictions with this test, a rare extra step in collections-based studies, allows us to identify ways in which the model's out-of-sample predictive ability could be improved. Lack of information on local variability in symbiont prevalence may simply be a feature of data derived from herbarium specimens. Natural history collectors sample one or a few specimens from local populations, and these observations are aggregated by the model to derive broad-scale estimates. This suggests that increasing local replication should be a factor considered in future collection efforts of natural history specimens, balancing the required time and effort along with limitations on storage space within collections. Herbarium collections were predominately used for taxonomic research in the past, but use of specimens to understand ongoing global change would benefit from increased collection efforts and expansion of herbarium collections. An alternative validation test would be to hold out samples from the historic data set. Such a test would more clearly match the conditions of the training data (i.e., in spatial scale and climate conditions); however, the trade-off between training and testing the model with a limited number of sampled specimens held us back from exploring this option. Splitting datasets can negatively impact model estimates, and the choice of how to split the data for model validation is not trivial (Bergmeir and Benítez 2012; James et al. 2013).

Another key consideration in forecasting the dynamics of host-microbe symbioses is the spatial scale of both specimen georeferencing and available climate data. For this analysis, most specimen localities were assigned coordinates at county or city centroids, and the climate data examined was on 4 km grid cells. Georeferencing of specimens as accurately as possible is a key

priority of herbarium specimen digitization efforts (Davis 2023; Soltis 2017). While the INLA modeling approach that we used allows for predictions at arbitrarily small spatial scales and would simplify connecting model predictions to the scale of a given climate driver, the coarse scale inherent to our analysis may obscure some local-scale ecological processes. Poor predictive ability at local scales in this grass-endophyte system is not surprising, as previous studies have found that local variation (e.g., in soil conditions, in microclimate), even to the scale of hundreds of meters, can structure endophyte-host niches (Gundel et al. 2024; Kazenel et al. 2015). Local adaptation in either the host or symbiont to microclimate or soil conditions could cause populations to differ from broad regional trends. The choice of prior distributions for spatially varying random effects also impacts the model's flexibility to capture spatial trends. Our exploration of model sensitivity to prior choice (presented in the *Supplemental Methods*) reveals qualitatively similar results across a broad range of priors. An important next step would be integrating data from local and regional scales through modeling to constrain estimates of local and regional variation.

Predicting future niche-shifts of hosts and symbionts will require considering the coupled dynamics of host-symbiont dispersal in addition to fitness benefits. For example, transplanting symbiotic and nonsymbiotic plants beyond the range edge of *A. hyemalis* could tell us whether low endophyte prevalence in that area (Figure 4a) is a result of environmental conditions that lead the symbiosis to have negative fitness consequences, or is a result of some historical contingency or dispersal limitation that has thus far limited the presence of symbiotic hosts from a region where they would otherwise flourish and provide resilience. Incorporating available climatic and soil layers as covariates is another obvious step that could improve predictions. These steps will bridge gaps that often exist between large but broad bioclimatic and biodiversity data and small but high-resolution data on biotic interactions, and move towards the goal of predicting the dynamics of microbial symbioses under climate change (Isaac et al. 2020; Miller et al. 2019).

4.3 | Herbaria for Global Change Research

Our analysis advances the use of herbarium specimens in *global change biology* in two ways. First and foremost, this is one of a growing number of studies to examine microbial symbiosis using specimens from natural history collections, and the first, to our knowledge, to link long-term changes in symbioses to changes in climate. The responses of microbial symbioses are a rich target for future studies within historic specimens, particularly those that take advantage of advances in sequencing technology. While we used relatively coarse presence/absence data based on fungal morphology, other studies have examined historic plant microbiomes using molecular sequencing and sophisticated bioinformatics techniques, but these studies have so far been limited to relatively few specimens at limited spatial extents (Bearchell et al. 2005; Bieker et al. 2020; Bradshaw et al. 2021, 2023; Gross et al. 2021; Heberling and Burke 2019; Yoshida et al. 2015). Much of this work highlights the important role that historic specimens can play in tracking pathogens, a particularly important area as climate change facilitates the spread of new diseases (Jean B. Ristaino 2020; Singh et al. 2023).

Continued advances in capturing historic DNA and in filtering out potential contamination during specimen storage (Bakker et al. 2020; Daru et al. 2019; Raxworthy and Smith 2021) will be imperative in the effort to scale up these efforts. This scaling up will be essential to be able to quantify changes not just in the prevalence of symbionts, but also in symbionts' intraspecific variation and evolutionary responses to climate change, as well as in changes in the wider host microbiome. With improved molecular insights from historic specimens, we could ask whether the broad increases in endophytes that we have identified reflect selection for particular genetic strains or chemotypes and how this selection varies across space. Answering these questions as well as the unknown questions that future researchers may ask also reiterates the value in capturing meta-information during ongoing digitization efforts at herbaria around the world and during the accession of newly collected specimens (Edwards et al. 2024; Lendemer et al. 2020).

The second major advance in this analysis is in accounting for several potential biases in the data observation process that may be common to many collections-based research questions by using a spatially explicit random effects model. Potential biases introduced by the sampling habits of collectors (Daru et al. 2018) and variation between contemporary researchers during the collection of trait data, if not corrected for, could lead to overconfident inference about the strength and direction of historic change (Figure 2). Previous studies that have quantified the effects of collector biases typically find them to be small (Davis et al. 2015; Meineke et al. 2019), and we similarly did not find that collector has a strong effect on the results of our analysis, but that scorer identity did impact results. It is difficult to distinguish whether the impact of scorers was driven by true differences in scorers' biases or by unintended spatial or temporal clustering of the specimens examined by each scorer (Clayton et al. 1993; Urdangarin et al. 2023). By underweighting endophyte-positive samples that are clustered spatially or by collector or observer, the endophyte prevalence model is appropriately accounting for nuisance variables and providing a conservative inference of endophyte change relative to the raw data. Spatial autocorrelation is another phenomenon likely common in data derived from herbarium specimens (Willems et al. 2022), which our spatially explicit analysis models among samples. Beyond spatial autocorrelation of outcomes, systematic differences in sampling across space can result in spatial bias.

One strength of herbaria as vehicles for global change research is the relative ease with which specimens from many distinct geographic locations can be examined. We visited just nine institutions in the central southern United States, and we were able to sample seeds from specimens across an area spanning over 300,000 sq. km, including specimens from Mexico and Canada. Despite this advantage, the specimens we examined are concentrated in the south-central United States, with fewer specimens in the rapidly warming northeastern United States reflecting the regional focus of herbaria. We provide a simulation analysis exploring the potential impact of spatially and temporally biased sampling (Appendix S1—Supporting Methods). We found that the spatially-varying coefficient model had a strong ability to recapitulate temporal trends across space in simulated data, and that this result was robust to relatively high levels of spatial

bias (80% of data missing from one spatial region). Simulation analyses that extend this work to consider the myriad ways herbarium data may be biased (i.e., testing different spatial arrangements and scales of spatial bias, or testing different sample sizes) would be extremely valuable (Daru et al. 2018; Erickson and Smith 2021; Gaul et al. 2020; Meineke and Daru 2021; Schmidt et al. 2025).

5 | Conclusion

Ultimately, a central goal of *global change biology* is to generate predictive insights into the future of natural systems on a rapidly changing planet. Beyond host-microbe symbioses, detecting ecological responses to anthropogenic global change and attributing their causes would inform public policy decision-makers and adaptive management strategies. Natural history specimens, such as the plant hosts examined in this study, have a clear role to play in informing global change biodiversity science, including building understanding of the dynamics of host-symbiont interactions (Davis 2023). This survey of historic endophyte prevalence is necessarily correlative, yet it serves as a foundation to develop better predictive models of the response of microbial symbioses to climate change. Combining the insights from this type of regional-scale survey with field experiments and physiological performance data could be invaluable to identify mechanisms driving shifts in host-symbiont dynamics. Evidence is strong that certain dimensions of climate change correlated with endophytes' temporal responses; however, we do not know why trends in prevalence were weak in some areas or how endophytes would respond to more extreme changes in climate. The "time machine" of natural history collections revealed evidence of mutualism resilience for grass-endophyte symbioses in the face of environmental change, but more extreme changes could potentially push one or both partners beyond their physiological limits, leading to the collapse of the mutualism; more research is needed to understand what those limits might be.

Author Contributions

Joshua C. Fowler: conceptualization, data curation, formal analysis, funding acquisition, methodology, visualization, writing – original draft, writing – review and editing. **Jacob Moutouama:** data curation, formal analysis, writing – review and editing. **Tom E. X. Miller:** conceptualization, data curation, formal analysis, funding acquisition, methodology, resources, supervision, writing – review and editing.

Acknowledgments

We thank Dr. Jessica Budke for help in drafting our initial destructive sampling plan, and to the many staff members of herbaria who facilitated our research visits, as well as to the hundreds of collectors who contributed to the natural history collections. Several high school and undergraduate researchers contributed to data collection, including A. Appio-Riley, P. Bilderback, E. Chong, K. Dickens, L. Dufresne, B. Gutierrez, A. Johnson, S. Linder, E. Scales, B. Scherick, K. Schrader, E. Segal, G. Singla, and M. Tucker. This research was supported by funding from the National Science Foundation Division of Environmental Biology (grants 1754468 and 2208857) and by funding from the Texas Ecolab Program. Five anonymous reviewers greatly improved earlier versions of this manuscript.

Conflicts of Interest

The authors declare no conflicts of interest.

Data Availability Statement

Data from this publication can be found through a publicly available repository (<https://doi.org/10.5061/dryad.rn8pk0pn0>). Code for analyses can be found through a publicly available repository (<https://github.com/joshuacfowler/EndoHerbarium>) that is permanently archived on Zenodo: DOI: 10.5281/zenodo.17298101. Code includes methods for downloading publicly available datasets, including temperature and precipitation data from the PRISM climate group (<https://prism.oregonstate.edu>) and GBIF occurrence records for *Epichloë* host species (*A. hyemalis*: <https://doi.org/10.15468/dl.ryqqcv>, *A. perennans*: <https://doi.org/10.15468/dl.wtne24>, *E. virginicus*: <https://doi.org/10.15468/dl.yt76uh>).

References

- Afkhami, M. E. 2012. "Fungal Endophyte–Grass Symbioses Are Rare in the California Floristic Province and Other Regions With Mediterranean-Influenced Climates." *Fungal Ecology* 5, no. 3: 345–352.
- Afkhami, M. E., P. J. McIntyre, and S. Y. Strauss. 2014. "Mutualist-Mediated Effects on Species' Range Limits Across Large Geographic Scales." *Ecology Letters* 17, no. 10: 1265–1273.
- Afkhami, M. E., and J. A. Rudgers. 2008. "Symbiosis Lost: Imperfect Vertical Transmission of Fungal Endophytes in Grasses." *American Naturalist* 172, no. 3: 405–416.
- Aitken, S. N., S. Yeaman, J. A. Holliday, T. Wang, and S. Curtis-McLane. 2008. "Adaptation, Migration or Extirpation: Climate Change Outcomes for Tree Populations." *Evolutionary Applications* 1, no. 1: 95–111.
- Ambrose, K. V., A. M. Koppenhöfer, and F. C. Belanger. 2014. "Horizontal Gene Transfer of a Bacterial Insect Toxin Gene Into the *Epichloë* Fungal Symbionts of Grasses." *Scientific Reports* 4, no. 1: 5562.
- Aslan, C. E., E. S. Zavaleta, B. Tershy, and D. Croll. 2013. "Mutualism Disruption Threatens Global Plant Biodiversity: A Systematic Review." *PLoS One* 8, no. 6: e66993.
- Bachl, F. E., F. Lindgren, D. L. Borchers, and J. B. Illian. 2019. "Inlabru: An R Package for Bayesian Spatial Modelling From Ecological Survey Data." *Methods in Ecology and Evolution* 10, no. 6: 760–766.
- Bacon, C. W., and J. F. White. 2018. "Stains, Media, and Procedures for Analyzing Endophytes." In *Biotechnology of Endophytic Fungi of Grasses*, 47–56. CRC Press.
- Bakka, H., H. Rue, G.-A. Fuglstad, et al. 2018. "Spatial Modeling With r-Inla: A Review." *Wiley Interdisciplinary Reviews: Computational Statistics* 10, no. 6: e1443.
- Bakker, F. T., V. C. Bieker, and M. D. Martin. 2020. "Herbarium Collection-Based Plant Evolutionary Genetics and Genomics." *Frontiers in Ecology and Evolution* 8: 603948.
- Bazely, D. R., J. P. Ball, M. Vicari, et al. 2007. "Broad-Scale Geographic Patterns in the Distribution of Vertically-Transmitted, Asexual Endophytes in Four Naturally-Occurring Grasses in Sweden." *Ecography* 30, no. 3: 367–374.
- Bearchell, S. J., B. A. Fraaije, M. W. Shaw, and B. D. L. Fitt. 2005. "Wheat Archive Links Long-Term Fungal Pathogen Population Dynamics to Air Pollution." *Proceedings of the National Academy of Sciences of the United States of America* 102, no. 15: 5438–5442.
- Beguín, J., S. Martino, H. Rue, and S. G. Cumming. 2012. "Hierarchical Analysis of Spatially Autocorrelated Ecological Data Using Integrated Nested Laplace Approximation." *Methods in Ecology and Evolution* 3, no. 5: 921–929.
- Berg, C. S., J. L. Brown, and J. J. Weber. 2019. "An Examination of Climate-Driven Flowering-Time Shifts at Large Spatial Scales Over 153 Years in a Common Weedy Annual." *American Journal of Botany* 106, no. 11: 1435–1443.
- Bergmeir, C., and J. M. Benítez. 2012. "On the Use of Cross-Validation for Time Series Predictor Evaluation." *Information Sciences* 191: 192–213.
- Bieker, V. C., F. Sánchez Barreiro, J. A. Rasmussen, M. Brunier, N. Wales, and M. D. Martin. 2020. "Metagenomic Analysis of Historical Herbarium Specimens Reveals a Postmortem Microbial Community." *Molecular Ecology Resources* 20, no. 5: 1206–1219.
- Blois, J. L., P. L. Zarnetske, M. C. Fitzpatrick, and S. Finnegan. 2013. "Climate Change and the Past, Present, and Future of Biotic Interactions." *Science* 341, no. 6145: 499–504.
- Bradshaw, M., U. Braun, M. Elliott, et al. 2021. "A Global Genetic Analysis of Herbarium Specimens Reveals the Invasion Dynamics of an Introduced Plant Pathogen." *Fungal Biology* 125, no. 8: 585–595.
- Bradshaw, M. J., J. Carey, M. Liu, et al. 2023. "Genetic Time Traveling: Sequencing Old Herbarium Specimens, Including the Oldest Herbarium Specimen Sequenced From Kingdom Fungi, Reveals the Population Structure of an Agriculturally Significant Rust." *New Phytologist* 237, no. 4: 1463–1473.
- Brem, D., and A. Leuchtman. 2001. "*Epichloë* Grass Endophytes Increase Herbivore Resistance in the Woodland Grass *Brachypodium sylvaticum*." *Oecologia* 126, no. 4: 522–530.
- Brem, D. O., and A. D. Leuchtman. 1999. "High Prevalence of Horizontal Transmission of the Fungal Endophyte *epichloë* Sylvatica." *Bulletin of the Geobotanical Institute ETH* 65, no. 3: 12.
- Card, S. D., M. J. Faville, W. R. Simpson, et al. 2014. "Mutualistic Fungal Endophytes in the Triticeae—Survey and Description." *FEMS Microbiology Ecology* 88, no. 1: 94–106.
- Cheng, S., Y.-N. Zou, K. Kuča, A. Hashem, E. F. Abd_Allah, and Q.-S. Wu. 2021. "Elucidating the Mechanisms Underlying Enhanced Drought Tolerance in Plants Mediated by Arbuscular Mycorrhizal Fungi." *Frontiers in Microbiology* 12: 4029.
- Clay, K., and C. Schardl. 2002. "Evolutionary Origins and Ecological Consequences of Endophyte Symbiosis With Grasses." *American Naturalist* 160, no. S4: S99–S127.
- Clayton, D. G., L. Bernardinelli, and C. Montomoli. 1993. "Spatial Correlation in Ecological Analysis." *International Journal of Epidemiology* 22, no. 6: 1193–1202.
- Côté, S. D., T. P. Rooney, J.-P. Tremblay, C. Dussault, and D. M. Waller. 2004. "Ecological Impacts of Deer Overabundance." *Annual Review of Ecology, Evolution, and Systematics* 35, no. 1: 113–147.
- Craven, K. D., P. T. W. Hsiau, A. Leuchtman, W. Hollin, and C. L. Schardl. 2001. "Multigene Phylogeny of *Epichloë* Species, Fungal Symbionts of Grasses." *Annals of the Missouri Botanical Garden* 88: 14–34.
- Crawford, K. M., J. M. Land, and J. A. Rudgers. 2010. "Fungal Endophytes of Native Grasses Decrease Insect Herbivore Preference and Performance." *Oecologia* 164: 431–444.
- Crossley, M. S., T. D. Meehan, M. D. Moran, J. Glassberg, W. E. Snyder, and A. K. Davis. 2022. "Opposing Global Change Drivers Counterbalance Trends in Breeding North American Monarch Butterflies." *Global Change Biology* 28, no. 15: 4726–4735.
- Daly, C., and K. Bryant. 2013. *The PRISM Climate and Weather System—An Introduction*, 2. PRISM climate group.
- Daru, B. H., E. A. Bowman, D. H. Pfister, and A. E. Arnold. 2019. "A Novel Proof of Concept for Capturing the Diversity of Endophytic Fungi Preserved in Herbarium Specimens." *Philosophical Transactions of the Royal Society B* 374, no. 1763: 20170395.
- Daru, B. H., D. S. Park, R. B. Primack, et al. 2018. "Widespread Sampling Biases in Herbaria Revealed From Large-Scale Digitization." *New Phytologist* 217, no. 2: 939–955.

- Davis, C. C., C. G. Willis, B. Connolly, C. Kelly, and A. M. Ellison. 2015. "Herbarium Records Are Reliable Sources of Phenological Change Driven by Climate and Provide Novel Insights Into Species' Phenological Cueing Mechanisms." *American Journal of Botany* 102, no. 10: 1599–1609.
- Davis, C. C. 2023. "The Herbarium of the Future." *Trends in Ecology & Evolution* 38, no. 5: 412–423.
- Davitt, A. J., C. Chen, and J. A. Rudgers. 2011. "Understanding Context-Dependency in Plant–Microbe Symbiosis: The Influence of Abiotic and Biotic Contexts on Host Fitness and the Rate of Symbiont Transmission." *Environmental and Experimental Botany* 71, no. 2: 137–145.
- Davitt, A. J., M. Stansberry, and H. A. Rudgers. 2010. "Do the Costs and Benefits of Fungal Endophyte Symbiosis Vary With Light Availability?" *New Phytologist* 188, no. 3: 824–834.
- Decunata, F. A., L. I. Pérez, D. P. Malinowski, M. A. Molina-Montenegro, and P. E. Gundel. 2021. "A Systematic Review on the Effects of *Epichloë* Fungal Endophytes on Drought Tolerance in Cool-Season Grasses." *Frontiers in Plant Science* 12: 644731.
- Di Luzio, M., G. L. Johnson, C. Daly, J. K. Eischeid, and J. G. Arnold. 2008. "Constructing Retrospective Gridded Daily Precipitation and Temperature Datasets for the Conterminous United States." *Journal of Applied Meteorology and Climatology* 47, no. 2: 475–497.
- Donald, M. L., T. F. Bohner, K. M. Kolis, R. A. Shadow, J. A. Rudgers, and T. E. X. Miller. 2021. "Context-Dependent Variability in the Population Prevalence and Individual Fitness Effects of Plant–Fungal Symbiosis." *Journal of Ecology* 109, no. 2: 847–859.
- Douglas, A. E. 1998. "Host Benefit and the Evolution of Specialization in Symbiosis." *Heredity* 81, no. 6: 599–603.
- Duan, Y.-W., H. Ren, T. Li, et al. 2019. "A Century of Pollination Success Revealed by Herbarium Specimens of Seed Pods." *New Phytologist* 224, no. 4: 1512–1517.
- Edwards, E. J., B. D. Mishler, and C. D. Davis. 2024. "University Herbaria Are Uniquely Important." *Trends in Plant Science* 29: 825–826.
- Engel, M., T. Mette, and W. Falk. 2022. "Spatial Species Distribution Models: Using Bayes Inference With INLA and SPDE to Improve the Tree Species Choice for Important European Tree Species." *Forest Ecology and Management* 507: 119983.
- Erickson, K. D., and A. B. Smith. 2021. "Accounting for Imperfect Detection in Data From Museums and Herbaria When Modeling Species Distributions: Combining and Contrasting Data-Level Versus Model-Level Bias Correction." *Ecography* 44, no. 9: 1341–1352.
- Evers, S. M., T. M. Knight, D. W. Inouye, et al. 2021. "Lagged and Dormant Season Climate Better Predict Plant Vital Rates Than Climate During the Growing Season." *Global Change Biology* 27, no. 9: 1927–1941.
- Fine, P. E. M. 1975. "Vectors and Vertical Transmission: An Epidemiologic Perspective." *Annals of the New York Academy of Sciences* 266, no. 1: 173–194.
- Fowler, J. C., M. L. Donald, J. L. Bronstein, and T. E. X. Miller. 2023. "The Geographic Footprint of Mutualism: How Mutualists Influence Species' Range Limits." *Ecological Monographs* 93, no. 1: e1558.
- Fowler, J. C., S. Ziegler, K. D. Whitney, J. A. Rudgers, and T. E. X. Miller. 2024. "Microbial Symbionts Buffer Hosts From the Demographic Costs of Environmental Stochasticity." *Ecology Letters* 27, no. 5: e14438.
- Frade, P. R., F. De Jongh, F. Vermeulen, J. Van Bleijswijk, and R. P. M. Bak. 2008. "Variation in Symbiont Distribution Between Closely Related Coral Species Over Large Depth Ranges." *Molecular Ecology* 17, no. 2: 691–703.
- Frederickson, M. E. 2017. "Mutualisms Are Not on the Verge of Breakdown." *Trends in Ecology & Evolution* 32, no. 10: 727–734.
- Gaul, W., D. Sadykova, H. J. White, et al. 2020. "Data Quantity Is More Important Than Its Spatial Bias for Predictive Species Distribution Modelling." *PeerJ* 8: e10411.
- GBIF.Org. 2025a. "Occurrence Download." <https://www.gbif.org/occurrence/download/0018831-250127130748423>.
- GBIF.Org. 2025b. "Occurrence Download." <https://www.gbif.org/occurrence/download/0018850-250127130748423>.
- GBIF.Org. 2025c. "Occurrence Download." <https://www.gbif.org/occurrence/download/0018851-250127130748423>.
- Gelman, A., and J. Hill. 2006. *Data Analysis Using Regression and Multilevel/Hierarchical Models*. Cambridge University Press.
- Gilman, S. E., M. C. Urban, J. Tewksbury, G. W. Gilchrist, and R. D. Holt. 2010. "A Framework for Community Interactions Under Climate Change." *Trends in Ecology & Evolution* 25, no. 6: 325–331.
- Granath, G., M. Vicari, D. R. Bazely, J. P. Ball, A. Puentes, and T. Rakocovic. 2007. "Variation in the Abundance of Fungal Endophytes in Fescue Grasses Along Altitudinal and Grazing Gradients." *Ecography* 30, no. 3: 422–430.
- Gross, A., C. Petitcollin, C. Dutech, et al. 2021. "Hidden Invasion and Niche Contraction Revealed by Herbaria Specimens in the Fungal Complex Causing Oak Powdery Mildew in Europe." *Biological Invasions* 23: 885–901.
- Gundel, P. E., L. A. Garibaldi, M. A. Martínez-Ghersa, and C. M. Ghersa. 2011. "Neotyphodium Endophyte Transmission to *loium multiflorum* Seeds Depends on the Host Plant Fitness." *Environmental and Experimental Botany* 71, no. 3: 359–366.
- Gundel, P. E., J. A. Rudgers, and C. M. Ghersa. 2011. "Incorporating the Process of Vertical Transmission Into Understanding of Host–Symbiont Dynamics." *Oikos* 120, no. 8: 1121–1128.
- Gundel, P. E., P. Sun, N. D. Charlton, C. A. Young, T. E. X. Miller, and J. A. Rudgers. 2020. "Simulated Folivory Increases Vertical Transmission of Fungal Endophytes That Deter Herbivores and Alter Tolerance to Herbivory in *poa autumnalis*." *Annals of Botany* 125, no. 6: 981–991.
- Gundel, P. E., A. C. Ueno, C. Casas, et al. 2024. "Temporal Host–Symbiont Dynamics in Community Contexts: Impacts of Host Fitness and Vertical Transmission Efficiency on Symbiosis Prevalence." *Functional Ecology* 38, no. 12: 2610–2622.
- Gundel, P. E., I. Zabalgogea, and B. R. V. De Aldana. 2011. "Interaction Between Plant Genotype and the Symbiosis With *Epichloë* Fungal Endophytes in Seeds of Red Fescue (*Festuca rubra*)." *Crop and Pasture Science* 62, no. 11: 1010–1016.
- Hart, E. K., and K. Bell. 2015. "Prism: Download Data From the Oregon Prism Project, R Package Version 0.0.6." <https://github.com/ropensci/prism>.
- Heberling, J. J., and D. J. Burke. 2019. "Utilizing Herbarium Specimens to Quantify Historical Mycorrhizal Communities." *Applications in Plant Sciences* 7, no. 4: e01223.
- Hijmans, R. J., S. Phillips, J. Leathwick, J. Elith, and R. J. Hijmans. 2017. "Package 'dismo'." *Circles* 9, no. 1: 1–68.
- HilleRisLambers, J., M. A. Harsch, A. K. Ettinger, K. R. Ford, and E. J. Theobald. 2013. "How Will Biotic Interactions Influence Climate Change–Induced Range Shifts?" *Annals of the New York Academy of Sciences* 1297, no. 1: 112–125.
- Iannone, L. J., J. F. White Jr, L. M. Giussani, D. Cabral, and M. V. Novas. 2011. "Diversity and Distribution of Neotyphodium-Infected Grasses in Argentina." *Mycological Progress* 10, no. 1: 9–19.
- IPCC. Climate change. 2021. "The Physical Science Basis, 2021." <https://www.ipcc.ch/report/ar6/wg1/>.
- Isaac, N., J. B., M. A. Jarzyna, P. Keil, et al. 2020. "Data Integration for Large-Scale Models of Species Distributions." *Trends in Ecology & Evolution* 35, no. 1: 56–67.
- James, G., D. Witten, T. Hastie, and R. Tibshirani. 2013. *An Introduction to Statistical Learning*. Vol. 112. Springer.

- Jiménez-Valverde, A. 2012. "Insights Into the Area Under the Receiver Operating Characteristic Curve (Auc) as a Discrimination Measure in Species Distribution Modelling." *Global Ecology and Biogeography* 21, no. 4: 498–507.
- Kahle, D., and H. Wickham. 2019. *Package 'ggmap'*. Vol. 5, 2021. CRAN.
- Kazenel, M. R., C. L. Debban, L. Ranelli, et al. 2015. "A Mutualistic Endophyte Alters the Niche Dimensions of Its Host Plant." *AoB Plants* 7: plv005.
- Knapp, R. A., G. M. Fellers, P. M. Kleeman, et al. 2016. "Large-Scale Recovery of an Endangered Amphibian Despite Ongoing Exposure to Multiple Stressors." *Proceedings of the National Academy of Sciences* 113, no. 42: 11889–11894.
- Kozlov, M. V., I. V. Sokolova, V. Zverev, A. A. Egorov, M. Y. Goncharov, and E. L. Zvereva. 2020. "Biases in Estimation of Insect Herbivory From Herbarium Specimens." *Scientific Reports* 10, no. 1: 12298.
- Lapeyrolerie, M., and C. Boettiger. 2023. "Limits to Ecological Forecasting: Estimating Uncertainty for Critical Transitions With Deep Learning." *Methods in Ecology and Evolution* 14, no. 3: 785–798.
- Lee, B. R., E. F. Alecrim, T. K. Miller, et al. 2024. "Phenological Mismatch Between Trees and Wildflowers: Reconciling Divergent Findings in Two Recent Analyses." *Journal of Ecology* 112, no. 6: 1184–1199.
- Lendemer, J., B. Thiers, A. K. Monfils, et al. 2020. "The Extended Specimen Network: A Strategy to Enhance Us Biodiversity Collections, Promote Research and Education." *Bioscience* 70, no. 1: 23–30.
- Leuchtmann, A. 1992. "Systematics, Distribution, and Host Specificity of Grass Endophytes." *Natural Toxins* 1, no. 3: 150–162.
- Leuchtmann, A., C. W. Bacon, C. L. Schardl, J. F. White Jr., and M. Tadych. 2014. "Nomenclatural Realignment of *Neotyphodium* Species With Genus *Epichloë*." *Mycologia* 106, no. 2: 202–215.
- Lindgren, F., H. Rue, and J. Lindström. 2011. "An Explicit Link Between Gaussian Fields and Gaussian Markov Random Fields: The Stochastic Partial Differential Equation Approach." *Journal of the Royal Statistical Society, Series B: Statistical Methodology* 73, no. 4: 423–498.
- Liu, C., P. M. Berry, T. P. Dawson, and R. G. Pearson. 2005. "Selecting Thresholds of Occurrence in the Prediction of Species Distributions." *Ecography* 28, no. 3: 385–393.
- Mc Cargo, D. D., L. J. Iannone, M. V. Vignale, C. L. Schardl, and M. S. Rossi. 2014. "Species Diversity of *Epichloë* Symbiotic With Two Grasses From Southern Argentinean Patagonia." *Mycologia* 106, no. 2: 339–352.
- McFall-Ngai, M., M. G. Hadfield, T. C. G. Bosch, et al. 2013. "Animals in a Bacterial World, a New Imperative for the Life Sciences." *Proceedings of the National Academy of Sciences* 110, no. 9: 3229–3236.
- Meehan, T. D., N. L. Michel, and H. Rue. 2019. "Spatial Modeling of Audubon Christmas Bird Counts Reveals Fine-Scale Patterns and Drivers of Relative Abundance Trends." *Ecosphere* 10, no. 4: e02707.
- Meijer, G., and A. Leuchtmann. 2000. "The Effects of Genetic and Environmental Factors on Disease Expression (Stroma Formation) and Plant Growth in *brachypodium sylvaticum* Infected by *Epichloë Sylvatica*." *Oikos* 91, no. 3: 446–458.
- Meineke, E. K., A. T. Classen, N. J. Sanders, and T. J. Davies. 2019. "Herbarium Specimens Reveal Increasing Herbivory Over the Past Century." *Journal of Ecology* 107, no. 1: 105–117.
- Meineke, E. K., and B. H. Daru. 2021. "Bias Assessments to Expand Research Harnessing Biological Collections." *Trends in Ecology & Evolution* 36, no. 12: 1071–1082.
- Meineke, E. K., C. C. Davis, and T. J. Davies. 2018. "The Unrealized Potential of Herbaria for Global Change Biology." *Ecological Monographs* 88, no. 4: 505–525.
- Meyer, A. R., M. Valentin, L. Liulevicius, et al. 2022. "Climate Warming Causes Photobiont Degradation and c Starvation in a Boreal Climate Sentinel Lichen." *American Journal of Botany* 110: e16114.
- Miller, D. A. W., K. Pacifici, J. S. Sanderlin, and B. J. Reich. 2019. "The Recent Past and Promising Future for Data Integration Methods to Estimate Species' Distributions." *Methods in Ecology and Evolution* 10, no. 1: 22–37.
- Park, D. S., I. Breckheimer, A. C. Williams, E. Law, A. M. Ellison, and C. C. Davis. 2019. "Herbarium Specimens Reveal Substantial and Unexpected Variation in Phenological Sensitivity Across the Eastern United States." *Philosophical Transactions of the Royal Society B* 374, no. 1763: 20170394.
- Parker, B. J., J. Hřeček, A. H. C. McLean, and H. C. J. Godfray. 2017. "Genotype Specificity Among Hosts, Pathogens, and Beneficial Microbes Influences the Strength of Symbiont-Mediated Protection." *Evolution* 71, no. 5: 1222–1231.
- Parniske, M. 2008. "Arbuscular Mycorrhiza: The Mother of Plant Root Endosymbioses." *Nature Reviews Microbiology* 6, no. 10: 763–775.
- Pauw, A., and J. A. Hawkins. 2011. "Reconstruction of Historical Pollination Rates Reveals Linked Declines of Pollinators and Plants." *Oikos* 120, no. 3: 344–349.
- Piao, S., Q. Liu, A. Chen, et al. 2019. "Plant Phenology and Global Climate Change: Current Progresses and Challenges." *Global Change Biology* 25, no. 6: 1922–1940.
- Poisot, T., G. Bergeron, K. Cazelles, et al. 2021. "Global Knowledge Gaps in Species Interaction Networks Data." *Journal of Biogeography* 48, no. 7: 1552–1563.
- Rafferty, N. E., P. J. CaraDonna, and J. L. Bronstein. 2015. "Phenological Shifts and the Fate of Mutualisms." *Oikos* 124, no. 1: 14–21.
- Ramampianandra, E. C., A. Scheidegger, J. Wydler, and N. Schuwirth. 2023. "A Comparison of Machine Learning and Statistical Species Distribution Models: Quantifying Overfitting Supports Model Interpretation." *Ecological Modelling* 481: 110353.
- Raxworthy, C. J., and B. T. Smith. 2021. "Mining Museums for Historical DNA: Advances and Challenges in Museumomics." *Trends in Ecology & Evolution* 36, no. 11: 1049–1060.
- Reno, F., I. Pons, and T. Hance. 2019. "Evolutionary Responses of Mutualistic Insect–Bacterial Symbioses in a World of Fluctuating Temperatures." *Current Opinion in Insect Science* 35: 20–26.
- Ristaino, J. B. 2002. "Tracking Historic Migrations of the Irish Potato Famine Pathogen, *phytophthora infestans*." *Microbes and Infection* 4, no. 13: 1369–1377.
- Ristaino, J. B. 2020. "The Importance of Mycological and Plant Herbaria in Tracking Plant Killers." *Frontiers in Ecology and Evolution* 7: 521.
- Ristaino, J. B., C. T. Groves, and G. R. Parra. 2001. "Pcr Amplification of the Irish Potato Famine Pathogen From Historic Specimens." *Nature* 411, no. 6838: 695–697.
- Roberts, E. L., and A. Ferraro. 2015. "Rhizosphere Microbiome Selection by *Epichloë* Endophytes of *Festuca arundinacea*." *Plant and Soil* 396: 229–239.
- Rodriguez, R. J., J. F. White Jr., A. E. Arnold, and A. R. Redman. 2009. "Fungal Endophytes: Diversity and Functional Roles." *New Phytologist* 182, no. 2: 314–330.
- Rolshausen, G., F. Dal Grande, A. D. Sadowska-Deś, J. Otte, and I. Schmitt. 2018. "Quantifying the Climatic Niche of Symbiont Partners in a Lichen Symbiosis Indicates Mutualist-Mediated Niche Expansions." *Ecography* 41, no. 8: 1380–1392.
- Rudgers, J. A., M. E. Afkhami, L. Bell-Dereske, et al. 2020. "Climate Disruption of Plant-Microbe Interactions." *Annual Review of Ecology, Evolution, and Systematics* 51, no. 1: 561–586.

- Rudgers, J. A., M. E. Afkhami, M. A. Rúa, A. J. Davitt, S. Hammer, and V. M. Huguet. 2009. "A Fungus Among Us: Broad Patterns of Endophyte Distribution in the Grasses." *Ecology* 90, no. 6: 1531–1539.
- Rudgers, J. A., and A. L. Swafford. 2009. "Benefits of a Fungal Endophyte in *Elymus virginicus* Decline Under Drought Stress." *Basic and Applied Ecology* 10, no. 1: 43–51.
- Rue, H., S. Martino, and N. Chopin. 2009. "Approximate Bayesian Inference for Latent Gaussian Models by Using Integrated Nested Laplace Approximations." *Journal of the Royal Statistical Society, Series B: Statistical Methodology* 71, no. 2: 319–392.
- Sage, R. F. 2020. "Global Change Biology: A Primer." *Global Change Biology* 26, no. 1: 3–30.
- Saikkonen, K., P. E. Gundel, and M. Helander. 2013. "Chemical Ecology Mediated by Fungal Endophytes in Grasses." *Journal of Chemical Ecology* 39: 962–968.
- Schardl, C. L., C. A. Young, J. R. Faulkner, S. Florea, and J. Pan. 2012. "Chemotypic Diversity of *Epichloae*, Fungal Symbionts of Grasses." *Fungal Ecology* 5, no. 3: 331–344.
- Schmidt, R. J., K. E. Saban, L. Struwe, and C. C. Davis. 2025. "The Collector Practices That Shape Spatial, Temporal, and Taxonomic Bias in Herbaria." In *New Phytologist*. Wiley.
- Semmartin, M., M. Omacini, P. E. Gundel, and I. M. Hernández-Agramonte. 2015. "Broad-Scale Variation of Fungal-Endophyte Incidence in Temperate Grasses." *Journal of Ecology* 103, no. 1: 184–190.
- Seneviratne, S. I., X. Zhang, M. Adnan, et al. 2021. "Weather and Climate Extreme Events in a Changing Climate." https://www.ipcc.ch/report/ar6/wg1/downloads/report/IPCC_AR6_WGI_Chapter11.pdf.
- Simpson, D., H. Rue, A. Riebler, T. G. Martins, and S. H. Sørbø. 2017. "Penalising Model Component Complexity: A Principled, Practical Approach to Constructing Priors." *Statistical Science* 32: 1–28.
- Singh, B. K., M. Delgado-Baquerizo, E. Egidi, et al. 2023. "Climate Change Impacts on Plant Pathogens, Food Security and Paths Forward." *Nature Reviews Microbiology* 21, no. 10: 640–656.
- Sneck, M. E., J. A. Rudgers, C. A. Young, and T. E. X. Miller. 2017. "Variation in the Prevalence and Transmission of Heritable Symbionts Across Host Populations in Heterogeneous Environments." *Microbial Ecology* 74: 640–653.
- Soltis, P. S. 2017. "Digitization of Herbaria Enables Novel Research." *American Journal of Botany* 104, no. 9: 1281–1284.
- Stocker, T. F., D. Qin, G.-K. Plattner, et al. 2013. "Technical Summary." In *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*, 33–115. Cambridge University Press.
- Sully, S., D. E. Burkepile, M. K. Donovan, G. Hodgson, and R. Van Woesik. 2019. "A Global Analysis of Coral Bleaching Over the Past Two Decades." *Nature Communications* 10, no. 1: 1–5.
- Tadych, M., M. S. Bergen, and J. F. White Jr. 2014. "Epichloë spp. Associated With Grasses: New Insights on Life Cycles, Dissemination and Evolution." *Mycologia* 106, no. 2: 181–201.
- Tian, Z., R. Wang, K. V. Ambrose, B. B. Clarke, and F. C. Belanger. 2017. "The Epichloë Festucae Antifungal Protein Has Activity Against the Plant Pathogen Sclerotinia Homoeocarpa, the Causal Agent of Dollar Spot Disease." *Scientific Reports* 7, no. 1: 5643.
- Tintjer, T., A. Leuchtman, and K. Clay. 2008. "Variation in Horizontal and Vertical Transmission of the Endophyte Epichloë Elymi Infecting the Grass *elymus hystrix*." *New Phytologist* 179, no. 1: 236–246.
- Toby Kiers, E., T. M. Palmer, A. R. Ives, J. F. Bruno, and J. L. Bronstein. 2010. "Mutualisms in a Changing World: An Evolutionary Perspective." *Ecology Letters* 13, no. 12: 1459–1474.
- Tredennick, A. T., G. Hooker, S. P. Ellner, and P. B. Adler. 2021. "A Practical Guide to Selecting Models for Exploration, Inference, and Prediction in Ecology." *Ecology* 102, no. 6: e03336.
- Treindl, A. D., J. Stapley, and A. Leuchtman. 2023. "Genetic Diversity and Population Structure of Epichloë Fungal Pathogens of Plants in Natural Ecosystems." *Frontiers in Ecology and Evolution* 11: 1129867.
- Truitt, A. M., M. Kapun, R. Kaur, and W. J. Miller. 2019. "Wolbachia Modifies Thermal Preference in *drosophila melanogaster*." *Environmental Microbiology* 21, no. 9: 3259–3268.
- Tuljapurkar, S. D. 1982. "Population Dynamics in Variable Environments. III. Evolutionary Dynamics of r-Selection." *Theoretical Population Biology* 21, no. 1: 141–165. [https://doi.org/10.1016/0040-5809\(82\)90010-7](https://doi.org/10.1016/0040-5809(82)90010-7).
- Urdangarin, A., T. Goicoa, and M. D. Ugarte. 2023. "Evaluating Recent Methods to Overcome Spatial Confounding." *Revista Matemática Complutense* 36, no. 2: 333–360.
- Vikuk, V., C. A. Young, S. T. Lee, et al. 2019. "Infection Rates and Alkaloid Patterns of Different Grass Species With Systemic Epichloë Endophytes." *Applied and Environmental Microbiology* 85, no. 17: e00465-19.
- von Cräutlein, M., M. Helander, H. Korpelainen, et al. 2021. "Genetic Diversity of the Symbiotic Fungus Epichloë Festucae in Naturally Occurring Host Grass Populations." *Frontiers in Microbiology* 12: 756991.
- Wang, Z., C. Li, and J. White. 2020. "Effects of Epichloë Endophyte Infection on Growth, Physiological Properties and Seed Germination of Wild Barley Under Saline Conditions." *Journal of Agronomy and Crop Science* 206, no. 1: 43–51.
- Ward, E. J., E. E. Holmes, J. T. Thorson, and B. Collen. 2014. "Complexity Is Costly: A Meta-Analysis of Parametric and Non-Parametric Methods for Short-Term Population Forecasting." *Oikos* 123, no. 6: 652–661.
- Warren, R. J., and M. A. Bradford. 2014. "Mutualism Fails When Climate Response Differs Between Interacting Species." *Global Change Biology* 20, no. 2: 466–474.
- Webster, N. S., R. E. Cobb, and A. P. Negri. 2008. "Temperature Thresholds for Bacterial Symbiosis With a Sponge." *ISME Journal* 2, no. 8: 830–842.
- White, J. F., and G. T. Cole. 1985. "Endophyte-Host Associations in Forage Grasses. i. Distribution of Fungal Endophytes in Some Species of *Lolium* and *Festuca*." *Mycologia* 77, no. 2: 323–327.
- Willems, F. M., J. F. Scheepens, and O. Bossdorf. 2022. "Forest Wildflowers Bloom Earlier as Europe Warms: Lessons From Herbaria and Spatial Modelling." *New Phytologist* 235, no. 1: 52–65.
- Willis, C. G., E. R. Ellwood, R. B. Primack, et al. 2017. "Old Plants, New Tricks: Phenological Research Using Herbarium Specimens." *Trends in Ecology & Evolution* 32, no. 7: 531–546.
- Xia, C., N. Li, Y. Zhang, C. Li, X. Zhang, and Z. Nan. 2018. "Role of Epichloë Endophytes in Defense Responses of Cool-Season Grasses to Pathogens: A Review." *Plant Disease* 102, no. 11: 2061–2073.
- Yoshida, K., E. Sasaki, and S. Kamoun. 2015. "Computational Analyses of Ancient Pathogen DNA From Herbarium Samples: Challenges and Prospects." *Frontiers in Plant Science* 6: 771.
- Yoshida, K., V. J. Schuenemann, L. M. Cano, et al. 2013. "The Rise and Fall of the *phytophthora infestans* Lineage That Triggered the Irish Potato Famine." *eLife* 2: e00731.

Supporting Information

Additional supporting information can be found online in the Supporting Information section. **Appendix S1:** gcb70577-sup-0001-AppendixS1.pdf.